

SISG 2022: Module 11  
Session 9: Bioethics and legal issues

1. The NIH awards research grants using tax-payer money. Because the public is essentially paying for the research, the NIH has stipulated that all data collected in these funded studies must be made available to other researchers and stored in a communal database, including genomic data. An Indigenous community has proposed a compelling genetic epidemiology research study, but does not want the genetic data deposited in the database for other researchers to access. The concern is that genetic data identifies specific individuals, that the community is identifiable, and that researchers accessing the genetic data may use it for research studies that the community does not approve and does not agree with.

<b>Bioethical category</b>	<b>Considerations</b>
Beneficence	
Non-maleficence	
Autonomy	
Justice	

- a. Use the table to map bioethical considerations for whether the NIH should still award this grant even if the genetic data are not deposited in the database. Consider at least two different stakeholder viewpoints.
- b. What other options could there be besides simply funding or not funding the research study?

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 Session 10: Rare variation

- 1) You just got a large grant to identify rare variants associated with type 2 diabetes. You have colleagues around the world that can give you access to DNA from their case-control studies. If you were to design a study to identify rare (allele frequency <1%) variants associated with type 2 diabetes, what approach would you take and why?
- High-depth whole genome sequencing
  - Low-depth whole genome sequencing
  - Whole exome sequencing
  - GWAS chip and imputation
  - Exome chip (custom array)

	<b>Advantage</b>	<b>Disadvantage</b>
<b>High-depth WGS</b>	can identify nearly all variants in the genome with high confidence	very expensive
<b>Low-depth WGS</b>	cost-effective and useful approach for association mapping	has limited accuracy for rare-variant identification and genotype calling; compared to deep sequencing, is subject to power loss if the same number of subjects is sequenced
<b>Whole-exome sequencing</b>	can identify all exonic variants; is less expensive than WGS	is limited to the exome
<b>GWAS chip and imputation</b>	inexpensive	has lower accuracy for imputed rare variants Will miss any variants unique to your sample
<b>Exome chip (custom array)</b>	much cheaper than exome sequencing	provides limited coverage for very rare variants and for non-Europeans is limited to target regions

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 Session 11: Gene x Environment Interactions

- 1) You are conducting a GxE interaction study, where the environmental exposure is smoking. Your colleagues have shared their data with you, which means you can include 25,050 subjects in your study! You need to harmonize the smoking variable across studies. The studies, their sample size and study-specific questions related to smoking can be found in the table. You are trying to build the biggest dataset you can, but you must be able to use the same definition of smoking. What are the samples sizes you could have in your study if you used the following definitions for your “smoking” exposure?
- Cigarettes per day
  - Ever smoker
  - Current smoker

(a)		
Study (N)	Smoking-related questions	Possible responses
Study 1 (2,500)	1. Do you currently smoke cigarettes?	Y/N
	2. If yes, how many cigarettes per day?	###
Study 2 (1,200)	1. Have you smoked more than 100 cigarettes in your lifetime?	Y/N
	2. If yes, do you currently smoke?	Y/N
	3. If yes, how many packs per day do you smoke?	###
Study 3 (8,500)	1. Have you ever smoked?	Y/N
Study 4 (1,250)	1. Do you currently smoke?	Y/N
Study 5 (4,200)	1. Do you smoke?	Y/N
	2. When did you first start smoking regularly?	Past year; 1–5 years ago; >5 years ago
Study 6 (6,600)	1. Have you smoked tobacco in the past month?	Y/N
Study 7 (800)	1. Have you ever smoked regularly?	Y/N
	2. If yes, do you still smoke?	Y/N
	3. If yes, how much do you smoke a day?	1–10 cigarettes, 11–20 cigarettes, 21–30 cigarettes, >30 cigarettes

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Session 12: Risk Prediction and Population Screening

- 1) Why would a polygenic risk score developed in a European ancestry cohort be unreliable for a person who does not have recent European ancestors? Hint: think about the mechanics of GWAS that give specific SNPs and loading values.
- 2) Discuss the ethical and social implications of using polygenic risk scores for embryo selection
  - a. How should OrchidHealth handle rapid scientific developments? What happens if after an embryo is selected, new research comes out that shows that high PRS for one disease is inversely related to another disease?

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Population Screening  
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- 3) Determine how whole population screening compares to cascade screening for cost-effectiveness. In the general US population, the collective variants causing Familial Hypercholesterolemia (FH) are found at a frequency of 1/250 (0.004). All first-degree relatives of a proband (patient) with FH have a 0.50 frequency of also having FH. Assume each genetic test costs \$250. Genotyping errors (leading to false positive test result) occur at a rate of 0.1%.
  - a. How much does it cost to detect one person with FH in the general population compared to among first degree relatives of a proband?
  - b. Consider the error rate of genotyping for this platform. How many false positives do you expect per true positives in the entire population compared to in cascade screening?
  - c. Variants in three genes are responsible for 80% of FH cases. What are strategies for identifying FH cases without variants in these three genes?

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Session 13: Mendelian randomization

- 1) Explore MR-Base (<http://www.mrbase.org>) to conduct your own MR study.
- 2) Run an MR study of body mass index and lung cancer risk following the example in class.