



Summer Institute  
in Statistical Genetics 2018

## Integrative Genomics

### 3b. Systems Biology and Epigenetics



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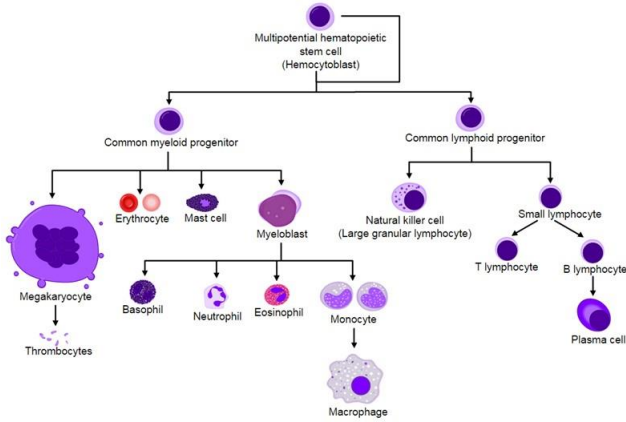
<http://www.gibsongroup.biology.gatech.edu>



#### Content of the Lecture

1. Immuno-Transcriptomics
2. Epigenome Projects from ENCODE to IHEC
3. Annotation of regulatory function
4. EpiWAS and the genetics of epigenome regulation

### Why Blood Gene Expression has such a high correlation structure



1. Because there are 3 common and dozens of rare blood cell types, and any cell-type biased gene expression correlates with abundance of the cell-type.
2. Because the environment, including disease status, modulates the expression of up to thousands of genes in a coordinated manner
3. The genetic component of most individual transcript abundance is regulated in trans, which also tends to lead to covariance – eg *Stat1* mediates the interferon response

Wikipedia: White Blood Cells

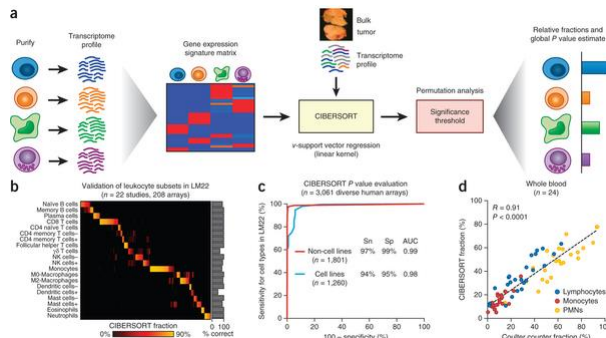
### CIBERSORT

Existing deconvolution methods perform accurately on distinct cell subsets in mixtures with well-defined composition (for example, blood), but are considerably less effective for discriminating closely related cell types (for example, naïve vs. memory B cells).

Input = reference gene expression signatures and unknown profile

Algorithm= linear support vector regression (SVR) – a machine learning approach robust to noise

Output = estimated abundances and p-value for the deconvolution

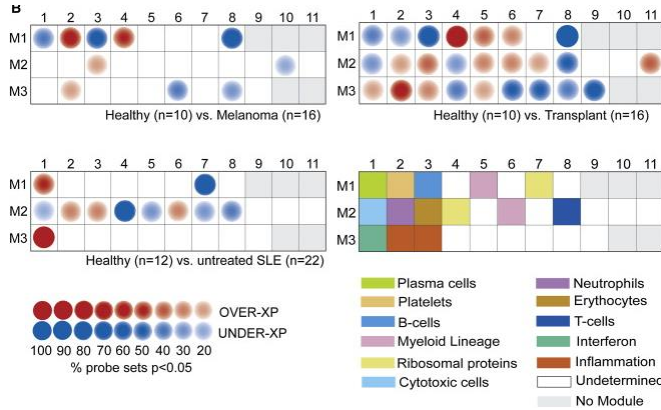


Newman et al (2015) *Nature Methods* 12: 453-457 “Robust enumeration of cell subsets from tissue expression profiles”

### Chaussabel Modules

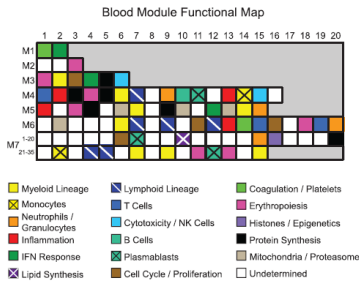
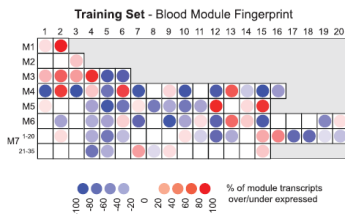
Used k-means clustering to search for conserved modules of genes that are differentially expressed in 8 diseases, namely 239 samples for SLE, JIA, T1D, melanoma, 2 types of bacteremia, influenza, or liver transplantation

Identified 28 modules involving 4742 transcripts (average of 170 per module)



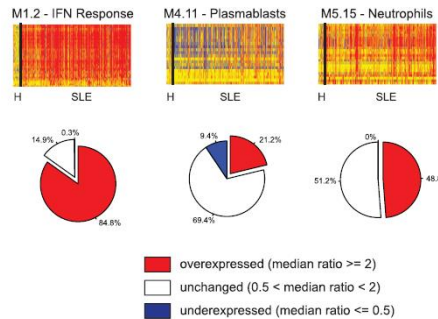
Chaussabel et al (2008) *Immunity* 29: 150-164 “A modular analysis framework for blood genomics studies: application to SLE”

### Update to 95 modules in 2016



158 Pediatric SLE patients  
924 longitudinal PB profiles (avg ~ 6 per patient)

First asked how modules correlate with disease, and how many patients show the effect

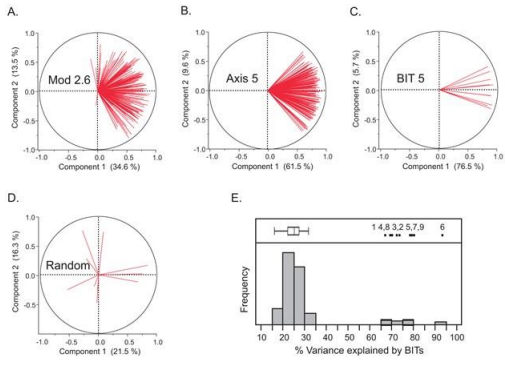


Banchereau et al (2016) *Cell* 165: 551-565 “Personalized Immunomonitoring Uncovers Molecular Networks that Stratify Lupus Patients”

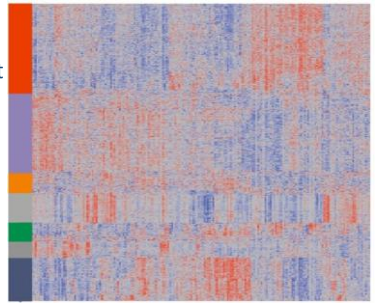
### Eight Axes capture basic Immune functions

We define the Axis scores at the first Principal Component of the positively correlated genes

Each Axis corresponds to an identified aspect of immune function, but they explain much more of the variance than the corresponding cell counts. The covariance is due to both cell abundance and transcription.



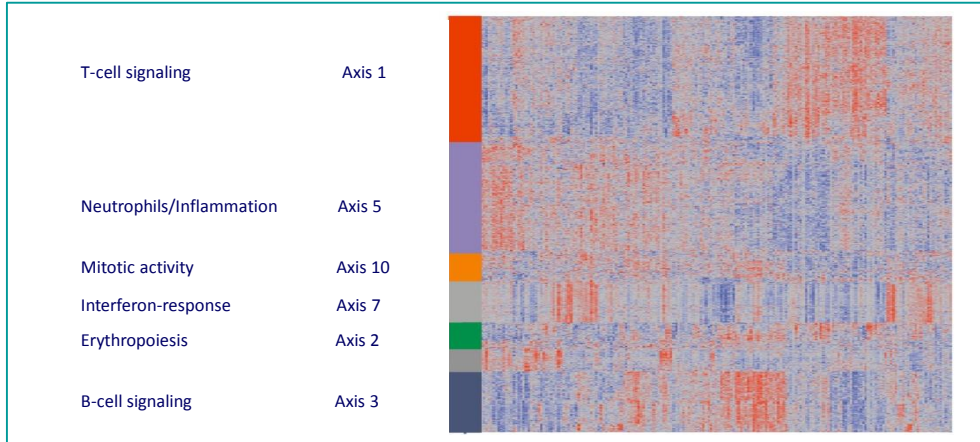
- Axis 1 T-cell signaling
- Axis 2 Reticulocyte development
- Axis 3 B-cell signaling
- Axis 4 Housekeeping functions
- Axis 5 Neutrophils and TLR
- Axis 6 Antibody response ?
- Axis 7 Interferon response
- Axis 10 Mitosis / cell cycle



BIT are 10 Blood Informative Transcripts that define each Axis.

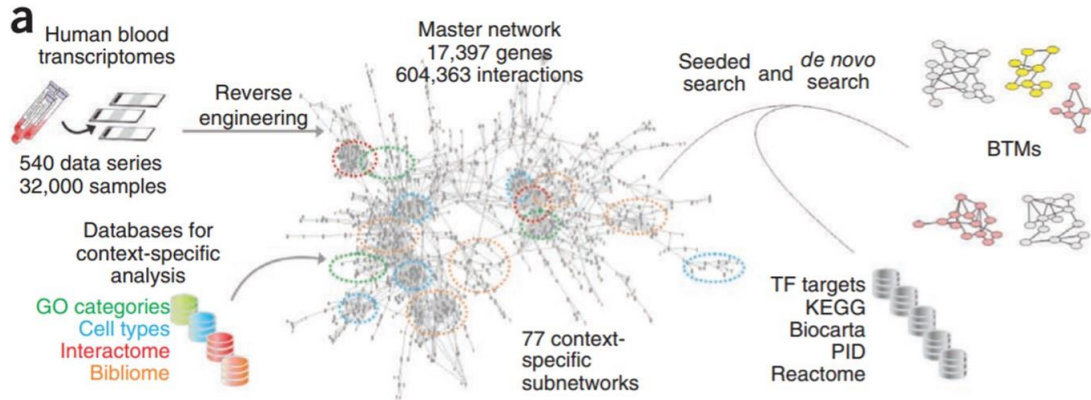
Preininger et al (2013) *PLoS Genet.* 9: e1003362 "Blood Informative Transcripts define nine common axes of peripheral blood gene expression"

← Individual samples →



↑ Individual transcripts ↓

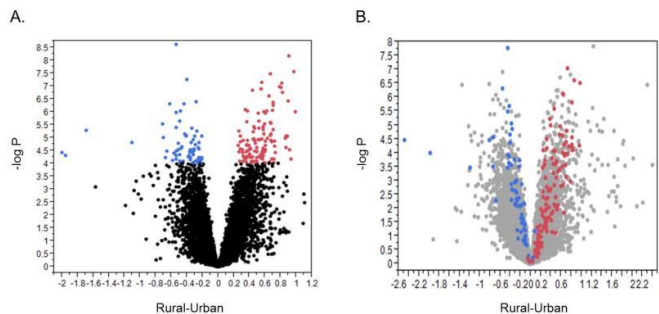
## Identifying Blood Transcript Modules



Li et al (2014) *Nat Immunology* 15: 195-204 “Molecular signatures of antibody responses derived from a systems biological study of 5 human vaccines”

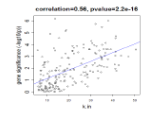
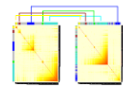
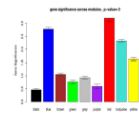
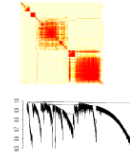
## The problem with gene ontology analysis on DE gene sets

1. Although powerful, DE analysis is also intrinsically under-powered, so there is a high false negative rate
2. Consequently, when you see a gene set annotated as “perturbed by drug x in cell-type y of females with disease z”, beware! Most likely a replicate of the experiment would give a completely different list.
3. Conversely, some annotations, eg “Lupus-associated genes” have multiple completely different lists.

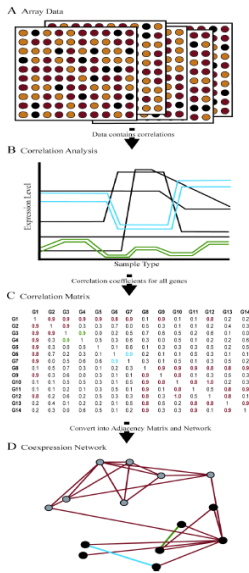


### Basic Workflow for Cluster analysis

1. Construct Similarity Matrix of Samples
2. Generate Modules with WGCNA (or MMC, or ...)
3. Perform Gene Ontology enrichment analysis on the Modules
4. Compare Module Preservation across datasets
5. Associate Module Eigenvectors with Traits OR search for Molecular Drivers of the Modules



### General Framework for Coexpression Network Analysis



1. Generate gene expression data (Microarray or RNASeq)
2. Measure Pearson correlations between all gene pairs
3. Dichotomize the matrix with some cutoff for the strength of correlation to generate an UNWEIGHTED adjacency matrix
4. OR Weight the correlations to generate a more nuanced network, for example using a power function:

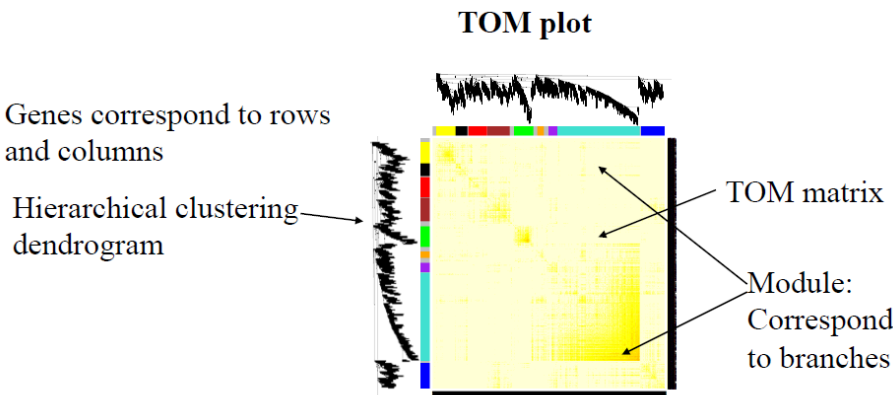
$$a_{ij} = |cor(x_i, x_j)|^\beta$$

Zhang and Horvath (2005) *SAGMB* 4: 17. A general framework for weighted gene co-expression network analysis.

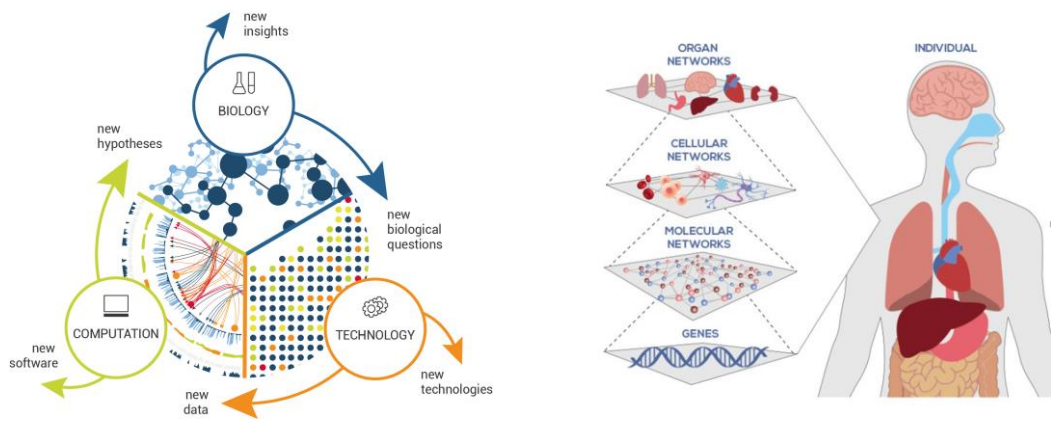
### Topological Overlap Matrices

Gene Modules correspond to Branches of the weighted hierarchical tree

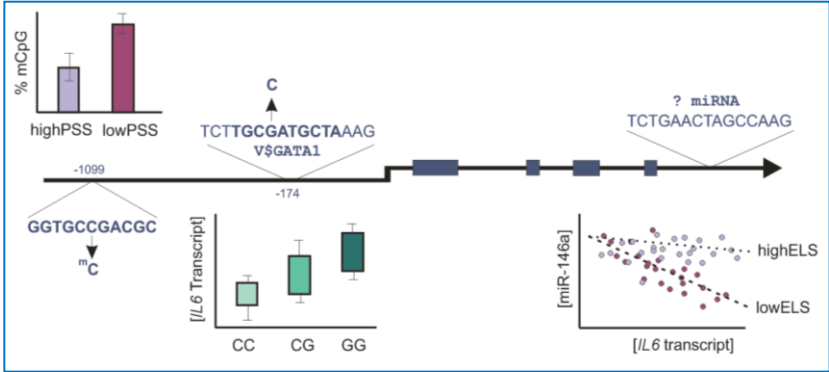
Each Modules is given a color – there may be dozens of them



### Integrative Systems Biology: big data meets cell biology



### The integrative nature of transcriptional regulation



<https://www.encodeproject.org/>

ENCODE: Encyclopedia of DNA Elements

The ENCODE (Encyclopedia of DNA Elements) Consortium is an international collaboration of research groups funded by the National Human Genome Research Institute (NHGRI). The goal of ENCODE is to build a comprehensive parts list of functional elements in the human genome, including elements that act at the protein and RNA levels, and regulatory elements that control cells and circumstances in which a gene is active.

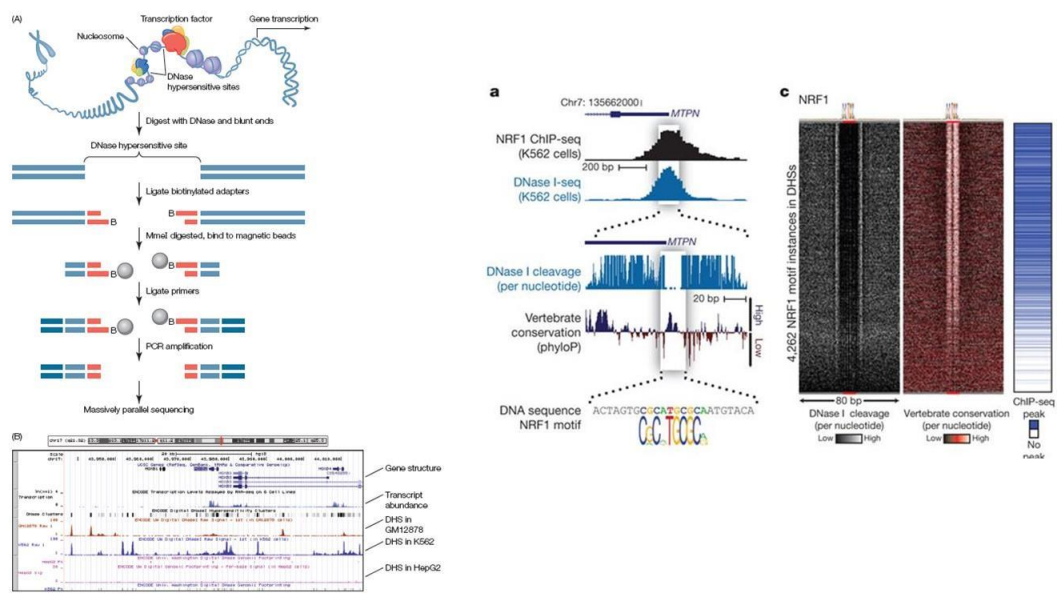
Project: 12876  
 Biosample Type: 12717  
 Assay Categories: ChIP-seq, RNA-seq, ATAC-seq, etc.

News: December releases: 48 ChIP-seq from the Reddy Lab

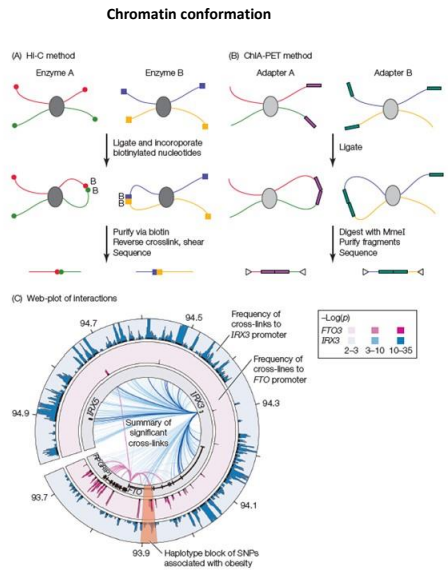
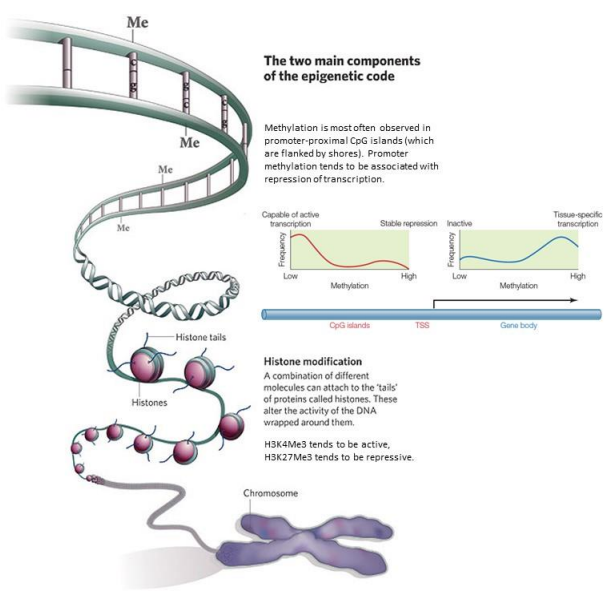
The ENCODE Project Consortium (2011) *PLOS Biology* 9: 1001046



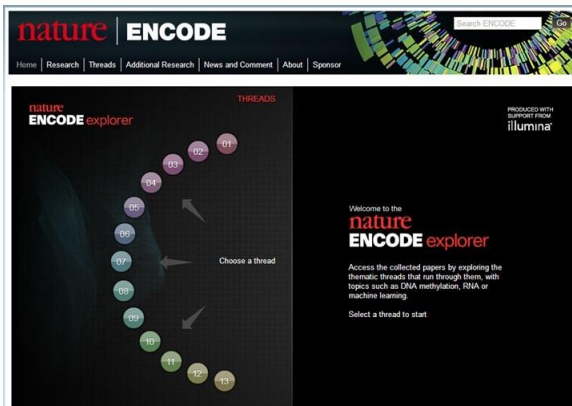
### DHS and TFBS: DNase hypersensitive sites and TF Binding



### Three modes of epigenetic regulation



## ENCODE Nature threads 2012



Thread	Topic
1	Transcription Factor Motifs
2	Chromatin patterns at Transcription Factor Binding Sites
3	Characterization of Intergenic Regions and Gene definition
4	RNA and Chromatin Modification patterns around Promoters
5	Epigenetic regulation of RNA Processing
6	Non-coding RNA characterization
7	DNA methylation
8	Enhancer discovery and characterization
9	Three-Dimensional connections across the Genome
10	Characterization of Network Topology
11	Machine Learning Approaches to Genomics
12	Impact of Functional Information on understanding Variation
13	Impact of Evolutionary Selection on functional regions

<http://www.nature.com/encode/#/threads>

## Roadmap Epigenomics Consortium

**Nature, 2015: INTEGRATIVE ANALYSIS OF 111 REFERENCE HUMAN EPIGENOMES**

Uniformly re-processed data, integrative analysis products and interactive browser sessions can be found at the supplementary website for the 2015 Consortium paper at <http://compbio.mit.edu/roadmap>

**DATA BROWSER**

The NIH Roadmap Epigenomics Mapping Consortium aims to produce a public resource of epigenomic maps for stem cells and primary ex vivo tissues selected to represent the normal counterparts of tissues and organ systems frequently involved in human disease.

**Data Releases**

The current release 9 of the Human Epigenome Atlas is a product of the NIH Roadmap Epigenomics Consortium. Release 9 contains a total of

**Download Data**

Data can be downloaded from the following sources:

- [Supplementary website for the 2015 Consortium paper](#)

<http://www.roadmapepigenomics.org/>

## Model Organism ENCODE

The screenshot shows the modENCODE website interface. At the top, it identifies itself as 'The National Human Genome Research Institute model organism ENCODEs of DNA Elements'. Navigation links include 'About modENCODE', 'Documentation', 'Contact Us', and 'Project Wiki'. A quote states: "The modENCODE Project will try to identify all of the sequence-based functional elements in the *Caenorhabditis elegans* and *Drosophila melanogaster* genomes."

Key features include:

- modMine**: A tool to explore hierarchical views of regulatory networks for fly and worm, with options for Networks, Regions, and Maps.
- amazon**: Access to the entire modENCODE data set available for analysis in the Amazon compute cloud.
- Dataset**: Find, view, and download datasets in bulk.
- FTP**: Download released data using the traditional FTP interface.

A section for 'Choose an organism below to see GBrowse, Dataset Search links.' lists various species including *C. elegans*, *C. breweri*, *C. briggsae*, *C. japonica*, *C. remanei*, *D. melanogaster*, *D. obscura*, *D. repleta*, *D. pseudoobscura*, *D. simulans*, *D. yakuba*, and *D. yoko*.

The 'Browse Projects' section is active for *D. melanogaster* and *C. elegans*. It lists several projects:

- Chromatin structure** (selected)
- Copy Number Variation
- Gene Structure
- Genome Sequence
- Histone modification and replacement
- Metadata only
- Other chromatin binding sites
- RNA expression profiles

Two specific projects are highlighted:

- Genome-wide Chromatin Profiling in *Drosophila***: Histone Variants. PI: Steven Henikoff, Labs: Kamran Ahmad. Submissions: 7. Experimental factors: biochemical fraction, cell line, extraction time, sodium chloride concentration. GBrowse Tracks: 2.
- Genome-wide Chromatin Profiling**: Histone Variants. PI: Steven Henikoff, Labs: Kamran Ahmad, Steven Henikoff. Submissions: 32. Experimental factors: developmental stage, biochemical fraction, cell line, extraction time, sodium chloride concentration. GBrowse Tracks: 32.

<http://www.modencode.org/>

## International Human Epigenome Consortium

The screenshot shows the IHEC website. The header includes the IHEC logo and navigation links: 'About', 'Research', 'IHEC Data Portal', 'News+Events', and 'Contact'.

The main banner features the text: "IHEC celebrates major coordinated paper release" and "Cell Press Special Edition International Human Epigenome Consortium Collection". Below the banner are navigation buttons for "Cell Papers", "BLUEPRINT Papers", "NIH Roadmap Papers", and "Annual Meeting".

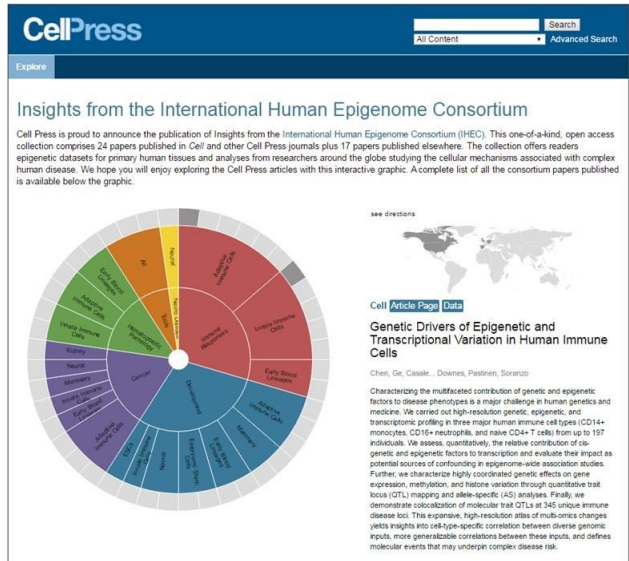
The 'Research' section includes a world map graphic and text: "By setting quality standards and providing efficient communication structures, IHEC fosters continuous exchange among scientists. It promotes rapid data sharing and minimizes redundancy between different individual research projects. Learn more about the research activities of IHEC."

The 'Why Epigenomics?' section features a family photo and text: "Epigenomics research and human health are closely linked to each other. Progress in this field of research will thus add to an improved understanding of diseases, and how to better treat and prevent them. Find out what makes epigenomics and the endeavor of IHEC so fascinating."

The 'IHEC Data Portal' section shows a grid of colored squares representing data points and text: "IHEC makes available comprehensive sets of reference epigenomes relevant to health and disease. You may view, search and download the data already released by the different IHEC-associated projects via the IHEC Data Portal."

<http://ihc-epigenomes.org/>

### IHEC Cell threads 2016



24 Papers published in Nov 2016 (Cell, Cell Reports, Cell Stem Cell, Cancer Cell)

<http://www.cell.com/consortium/IHEC>

### Enrichment of regulatory elements at GWAS loci

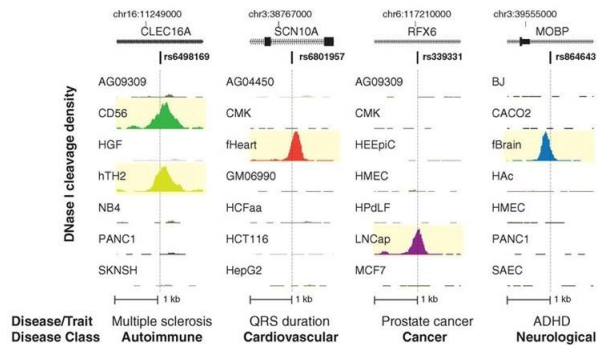
93% of GWAS peak SNPs are located in regulatory regions rather than affecting the protein sequence

Maurano et al performed DNase-Seq on 349 cell and tissue samples, identifying ~ 200,000 DHS per sample (2% of DNA)

75% of 5,130 GWAS peak SNPs are in a DHS, many specifically in a tissue expected to relate to pathology

419 of these pair with active promoters by Chia-PET, 40% acting over 250kb and 80% not with the closest gene

20% - 40% show allelic imbalance for chromatin accessibility



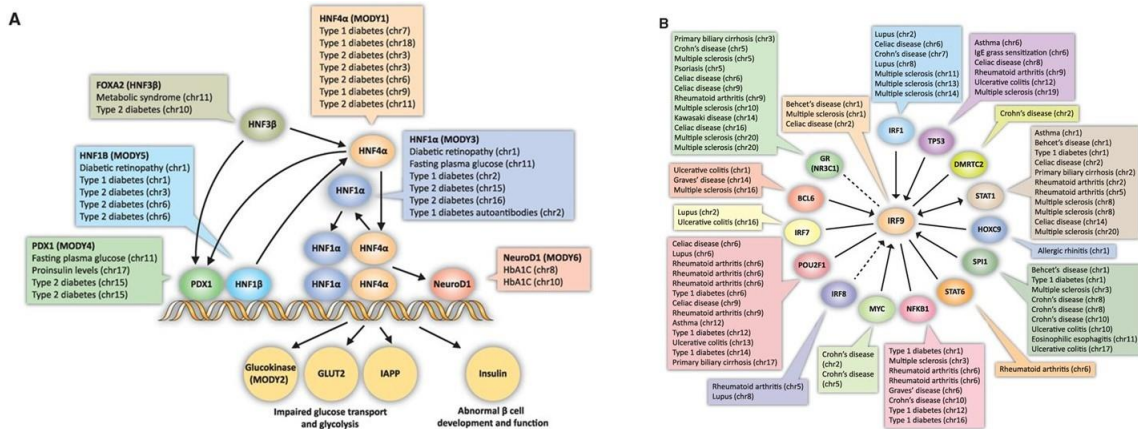
Maurano et al (2012) *Science* **337**: 1190-1195

### Disease associations cluster in regulatory pathways

(A) Monogenic diabetes locus TFBS are enriched at GWAS / DHS sites for Types 1 and 2 diabetes

(B) Transcription factors associated with multiple autoimmune diseases are enriched at GWAS / DHS sites

Similar results observed for several types of cancer and neurological disorders



Maurano et al (2012) *Science* **337**: 1190-1195

### CADD score annotation of likely deleteriousness

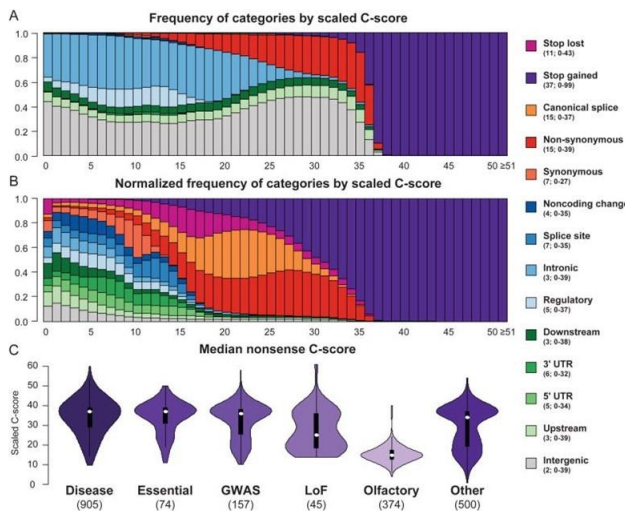
<http://cadd.gs.washington.edu/>

CADD (combined annotation dependent depletion) is an index from the Shendure lab at UW that summarizes evidence from 63 annotations encompassing:

- Functional or regulatory annotation
- Allele frequency and diversity
- Evolutionary conservation

The raw C-score is scaled to a relative CADD score as the  $-10 * \log_{10}(\text{rank}/\text{total})$ , namely:  
 30 is the top 0.1% of likely deleterious  
 20 is in the top 1%  
 10 is in the top 10%

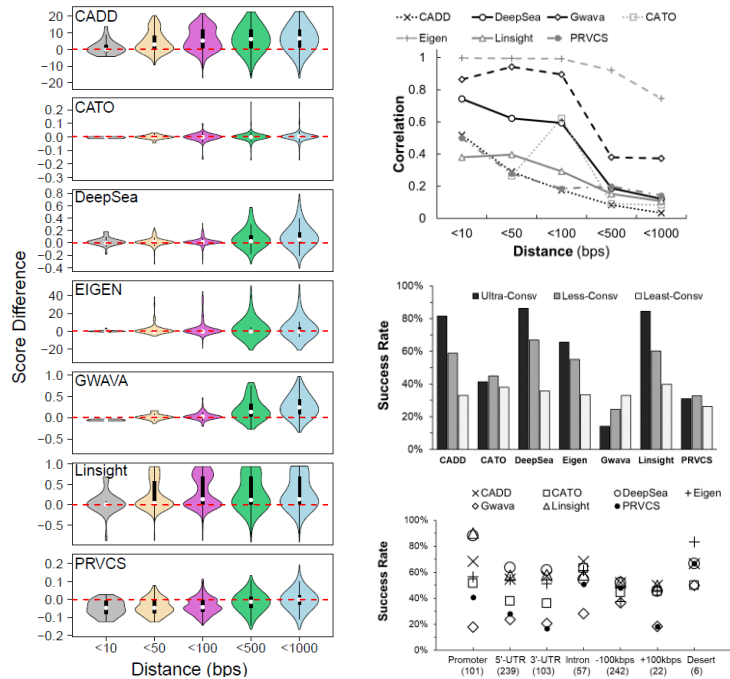
The score attempts unbiased prediction of "deleteriousness", based on machine learning comparison of 15M observed and simulated human variants



Kircher et al (2014) *Nature Genetics* **46**: 310-315

## Beware Regulatory Annotation

Li Liu, Max Sanderford, Sudhir Kumar, GG  
Under review



## Some (concise) definitions

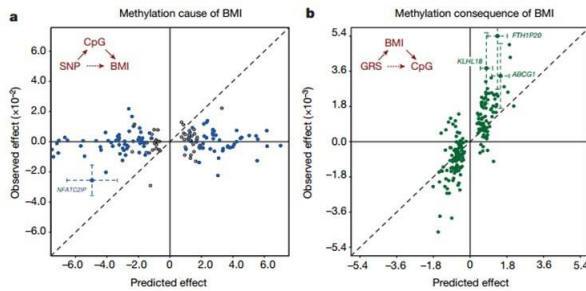
- GWAS: Genome-wide association study – search for SNPs significantly associated with a trait (eSNPs)
- TWAS: Transcriptome-wide association study – search for transcripts significantly associated with a trait (QTT)
- EpiWAS: Epigenome-wide association study – search for epigenetic marks significantly associated with a trait (EWAS also used, but earlier used to refer to Environment-wide association study)
- eQTL: a SNP which influences the abundance of a transcript. Cis-eQTL act locally (~ within  $\pm 500\text{kb}$ )
- eGene: a gene whose transcript abundance is regulated by a locally-acting SNP
- meQTL: a genotype which is associated with the degree of methylation at a CpG site
- Methyl  $\beta$ : typical measure of the degree of methylation, ranging from 0 to 1 (none to complete)
- hQTL: a genotype that is associated with the intensity of a histone mark (may be acetylation or methylation)
- ccQTL: a genotype that influences the level of chromatin conformation / cross-linking

## Epigenome-Wide Association Studies (EpiWAS) for Metabolic Disease

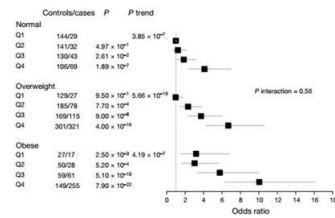
Methyl450 array study of whole blood DNA for 5,387 Europeans and Asians  
 Identified 278 CpG sites in 207 genes associated with BMI at  $p < 10^{-7}$ : consistent across ethnicities, 90% replicated

Similar effects observed in T cells and neutrophils in independent sample of 60 adults,  
 about half of the sites also associated with BMI in fat, liver, muscle

However, Mendelian randomization of SNPs that associate with both BMI and methylation level (meQTL)  
 implies that only a single site is causal – the majority are responsive to obesity  
 and in turn are explained by variation in blood glucose and lipids which may mediate the methylation



Methylation Risk Score predicts T2D somewhat independent of classical risk factors



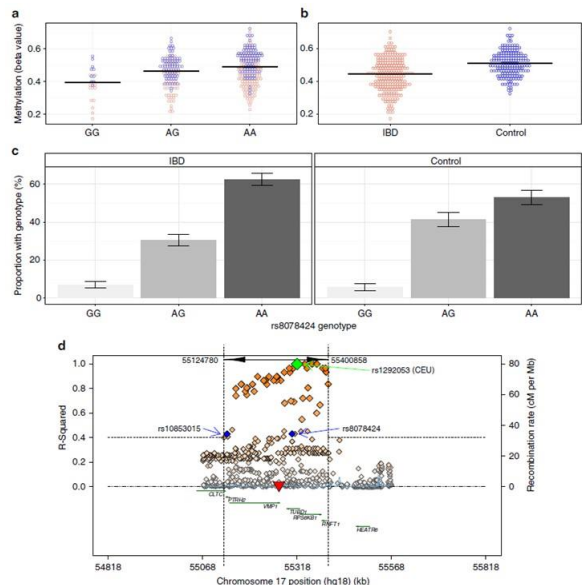
Wahl et al (2016) *Nature* 541: 81-85

## meQTL for Inflammatory Bowel Disease

VMP1 methylation is influenced by an meQTL, and associates with IBD

An meQTL SNP associates with IBD

Two meQTL SNPs are in mild LD with the GWAS SNP, and flank the CpG site



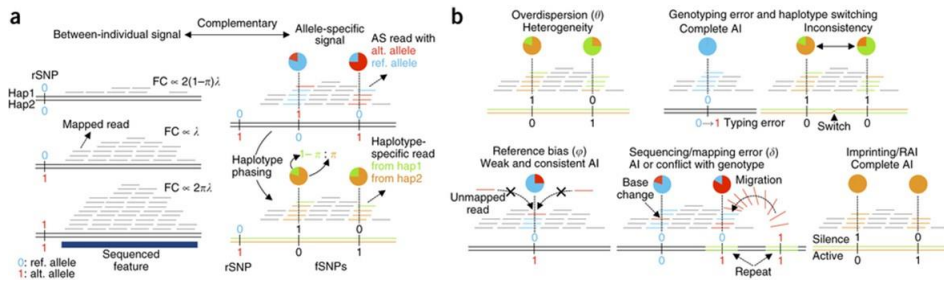
Ventham et al (2016) *Nature Communications* 7: 13507

### ATAC-Seq and enhancer detection

There are three basic approaches for detecting active chromatin, which is interpreted as enhancers:

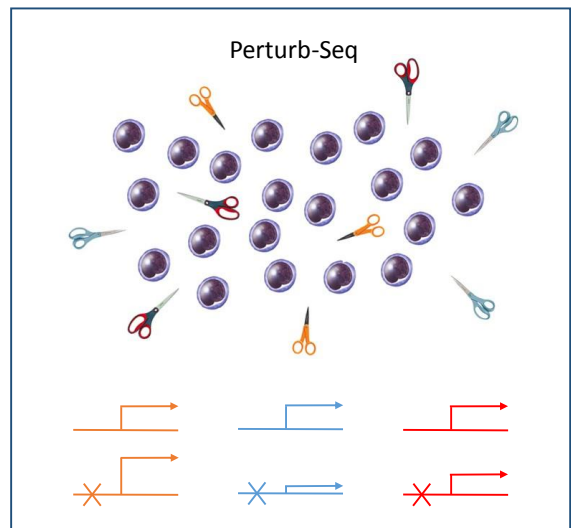
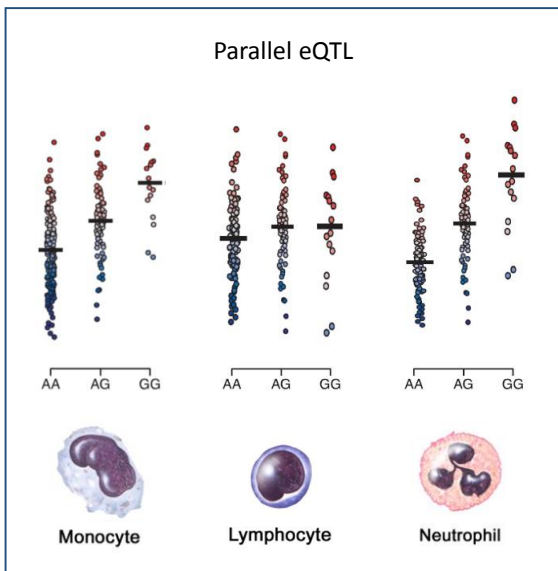
- DNase Hypersensitivity Site Sequencing (DNaseSeq)
- Chromatin immunoprecipitation Sequencing with CTCF, other TFs (ChIP-Seq)
- Assay for Transcriptionally Active Chromatin (ATAC-Seq)

An emerging software for allele-specific ATAC-Seq (and RNASeq) analysis is RASQUAL  
(Robust Allele-Specific Quantitation and Quality Control)



Kumasaka, Knights and Gaffney (2015) *Nature Genetics* 48: 206-13

### Single Cell Genetics



Adamson *et al* (2016) *Cell* 167: 1867-1882  
Datlinger *et al* (2017) *Nat Methods* 14: 297-301