Searching for Signatures of Selection with Selscan

Ryan Hernandez

The Effect of Positive Selection

Adaptive

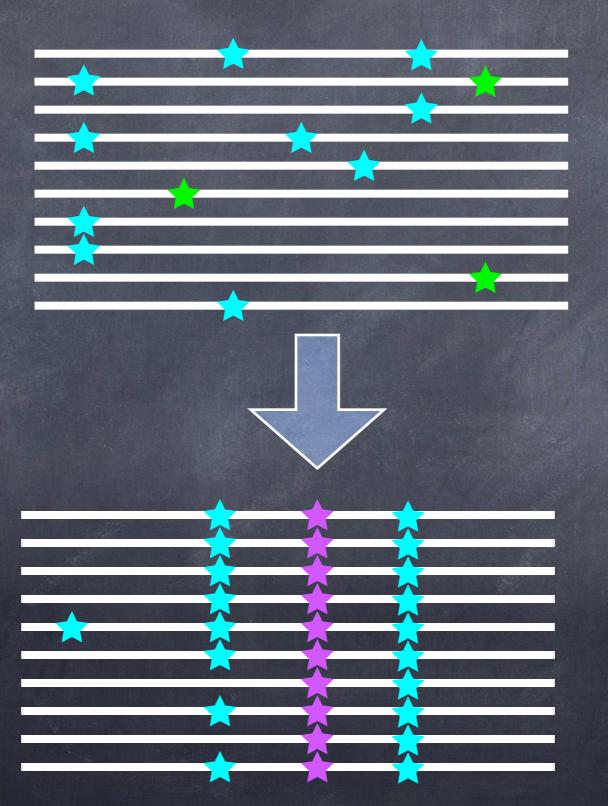
Neutral

Nearly Neutral

Mildly Deleterious

Fairly Deleterious

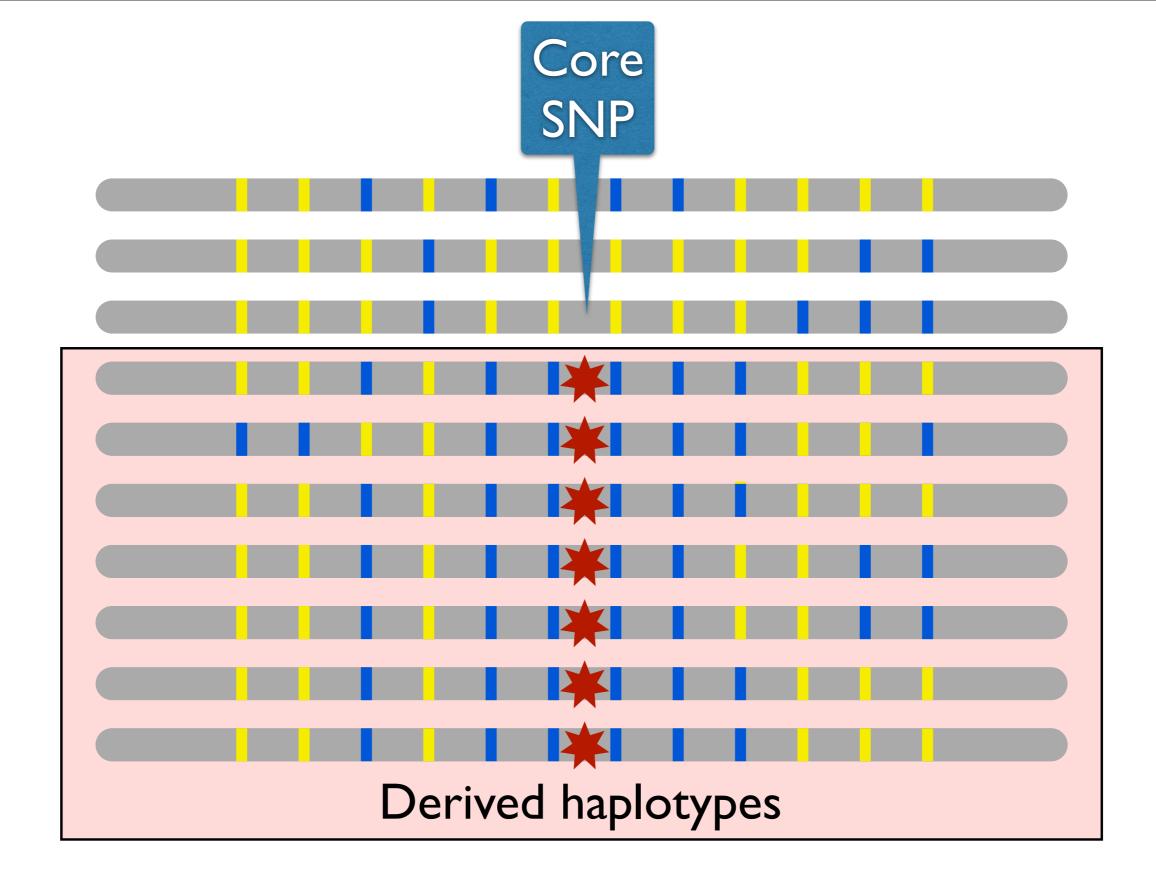
Strongly Deleterious

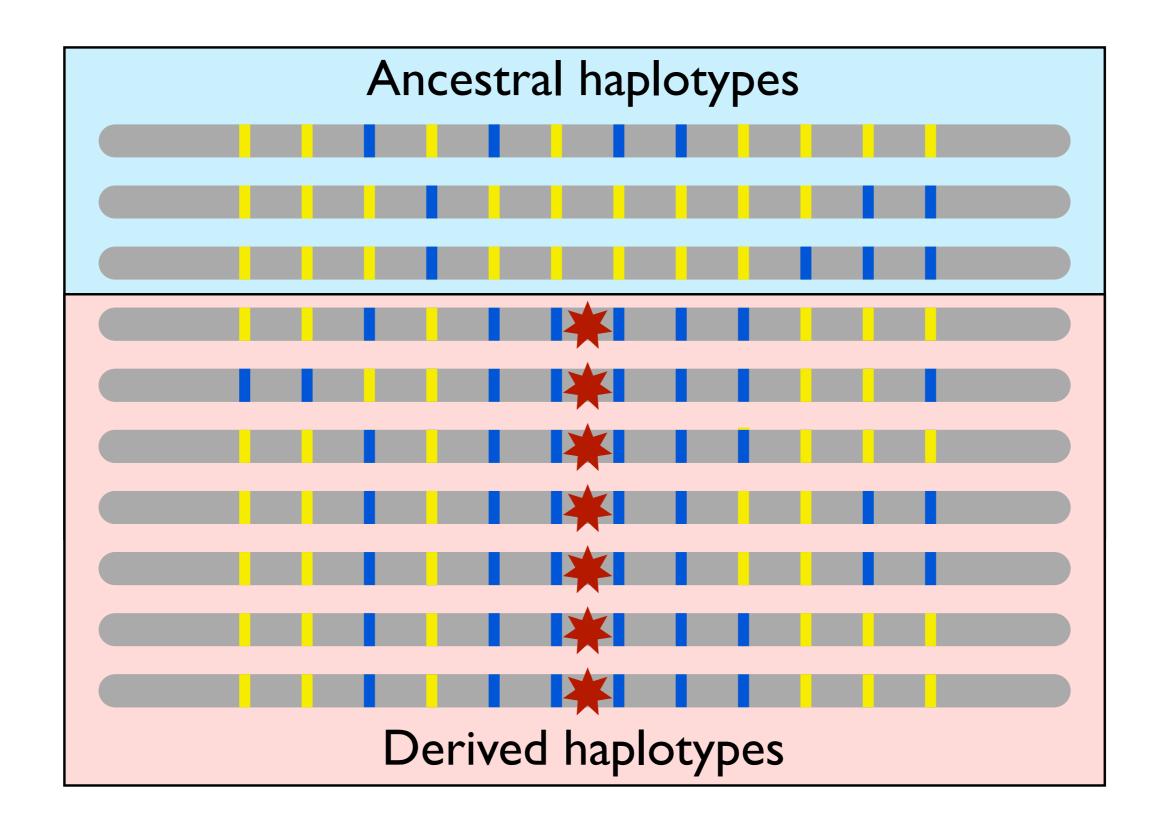


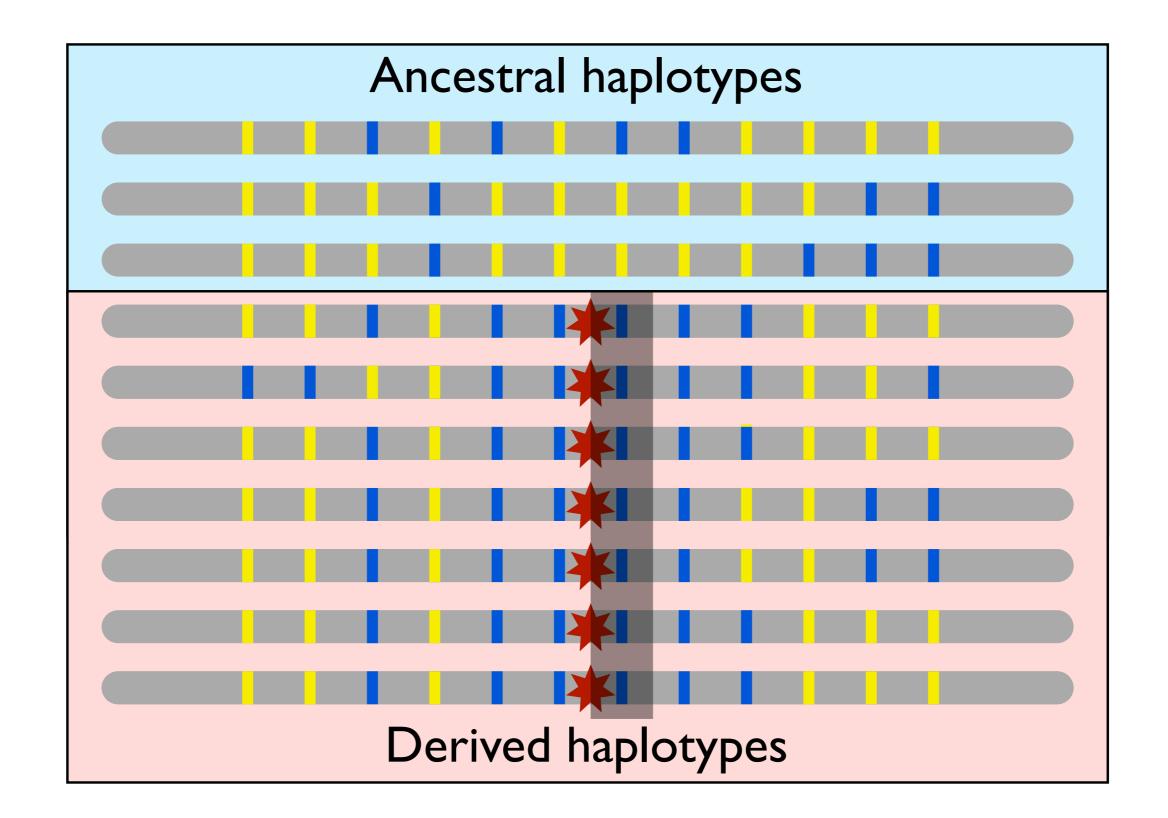
How do we capture this process in a statistic?

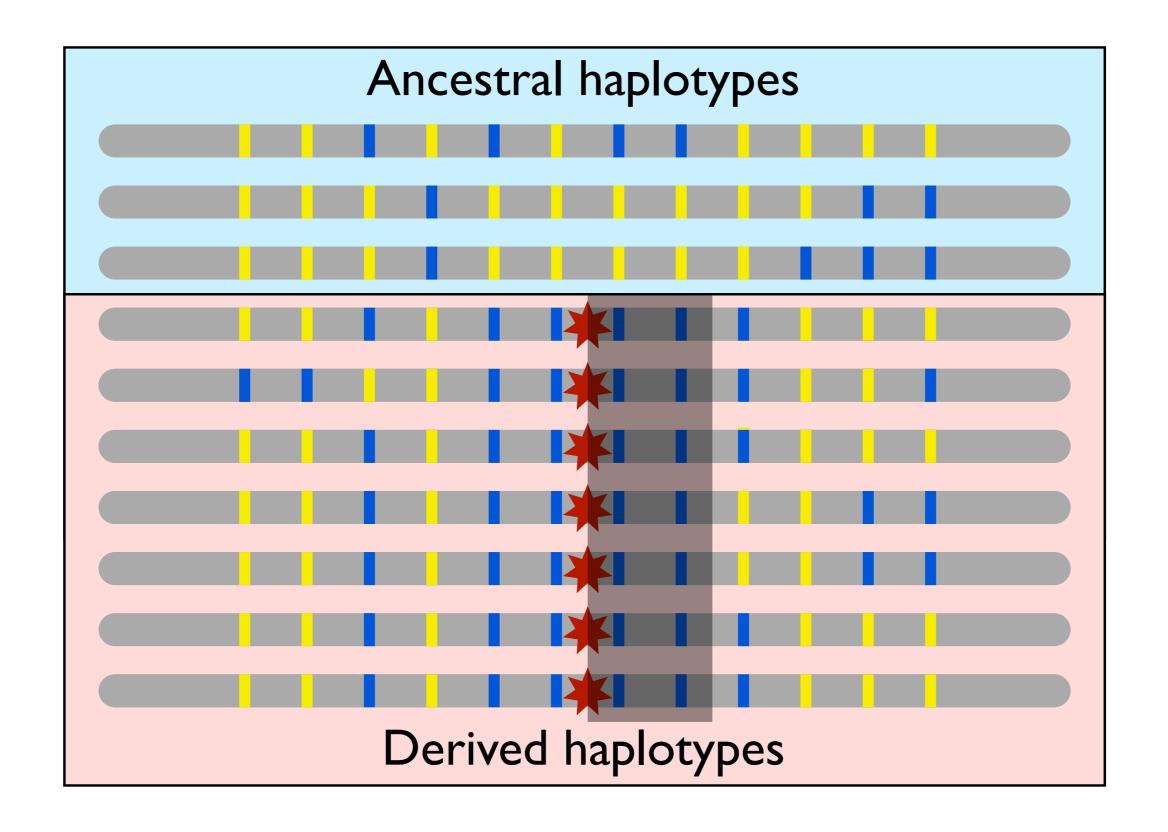
Extended Haplotype Homozygosity

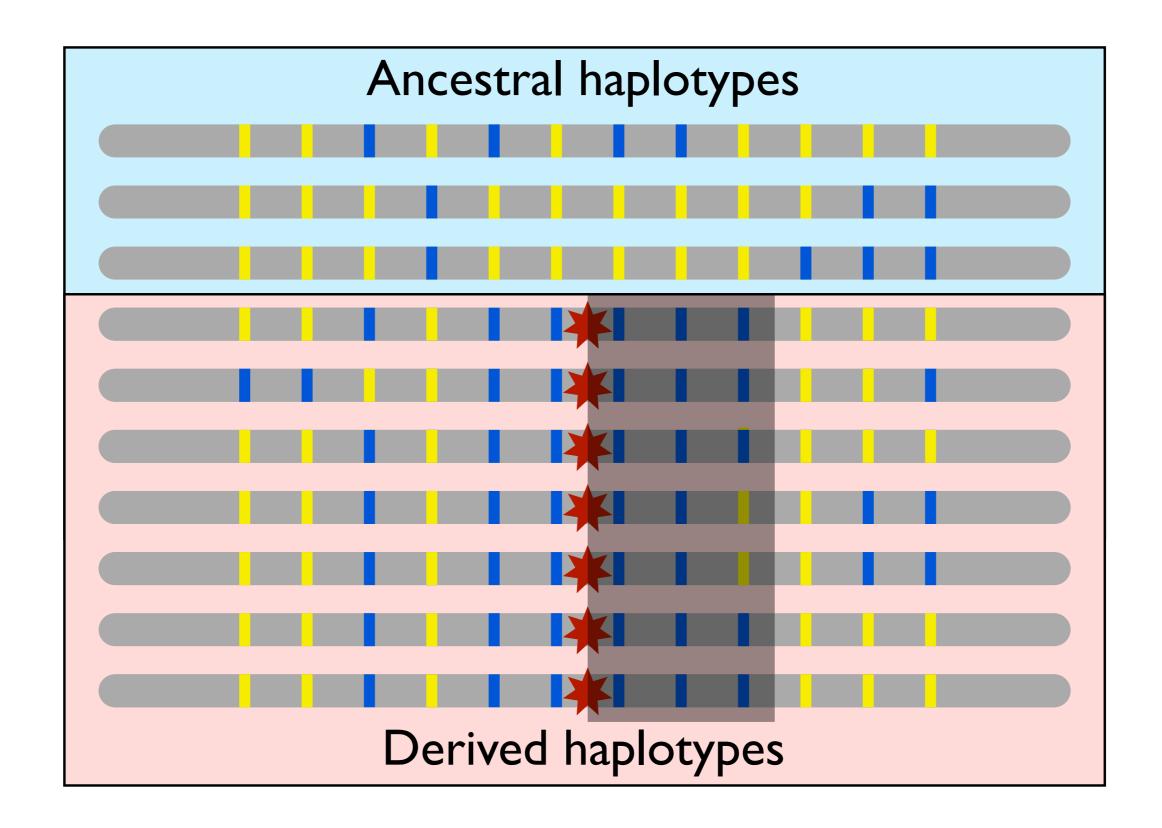
- Sabeti, et al. (Nature, 2002) proposed EHH
- Designed to track the decay of haplotype identity away from a locus of interest
 - If selection acts quickly enough
- Originally derives from ideas in Hudson, et al. (Genetics, 1994).

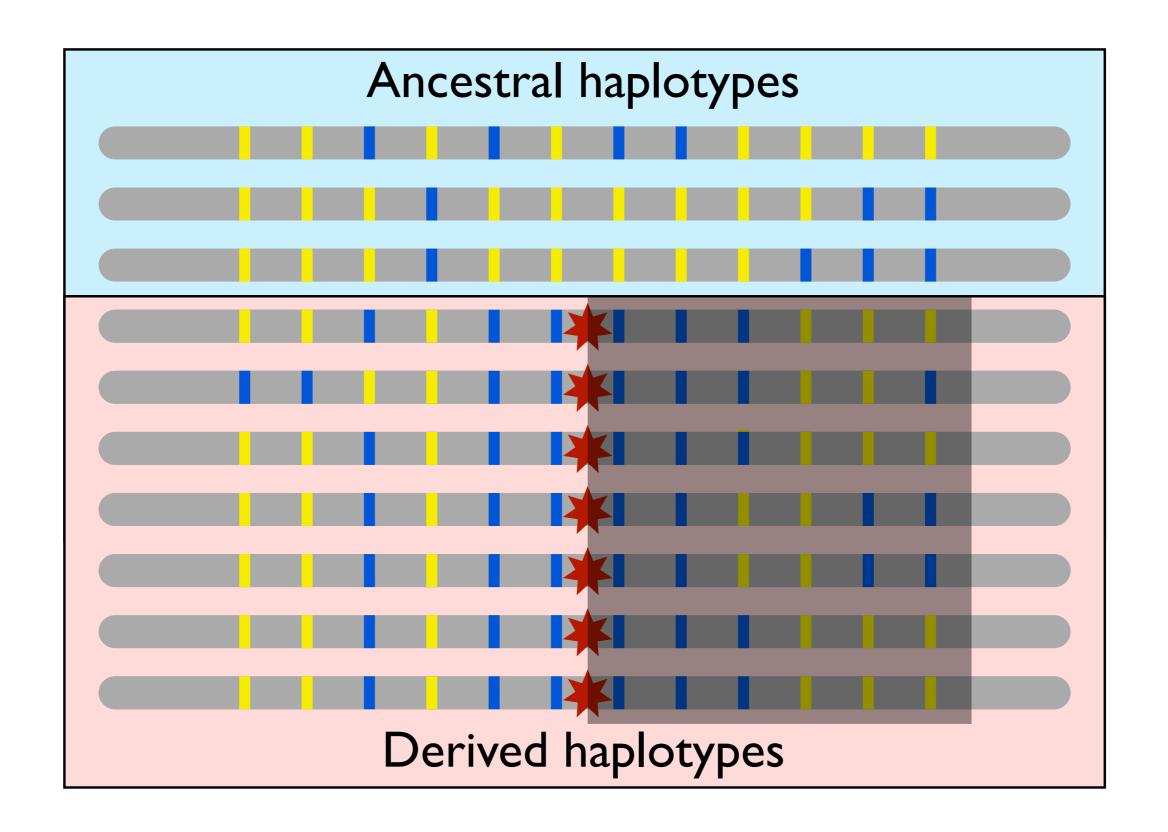












- ullet Given a locus of interest, $\mathcal C$ is the set of all distinct haplotypes at that locus.
- Select a "core" haplotype, $c \in C$.
- $\mathcal{H}(c,x)$ is the set of all distinct haplotypes that extend from the locus of interest to marker x and contain the core haplotype c.
- For $h \in \mathcal{H}(c,x)$, n_h is the number of haplotypes of type h
- \bullet n_c is the number of the core haplotypes

• If $EHH_c(x)$ is the extended haplotype homozygosity of the core haplotype c out to marker x, then

$$EHH_c(x) = \sum_{h \in \mathcal{H}(c,x)} \frac{\binom{n_h}{2}}{\binom{n_c}{2}}$$

$$\binom{n}{2} := 0 \ \forall n < 2$$

• If $EHH_c(x)$ is the extended haplotype homozygosity of the core haplotype c out to marker x, then

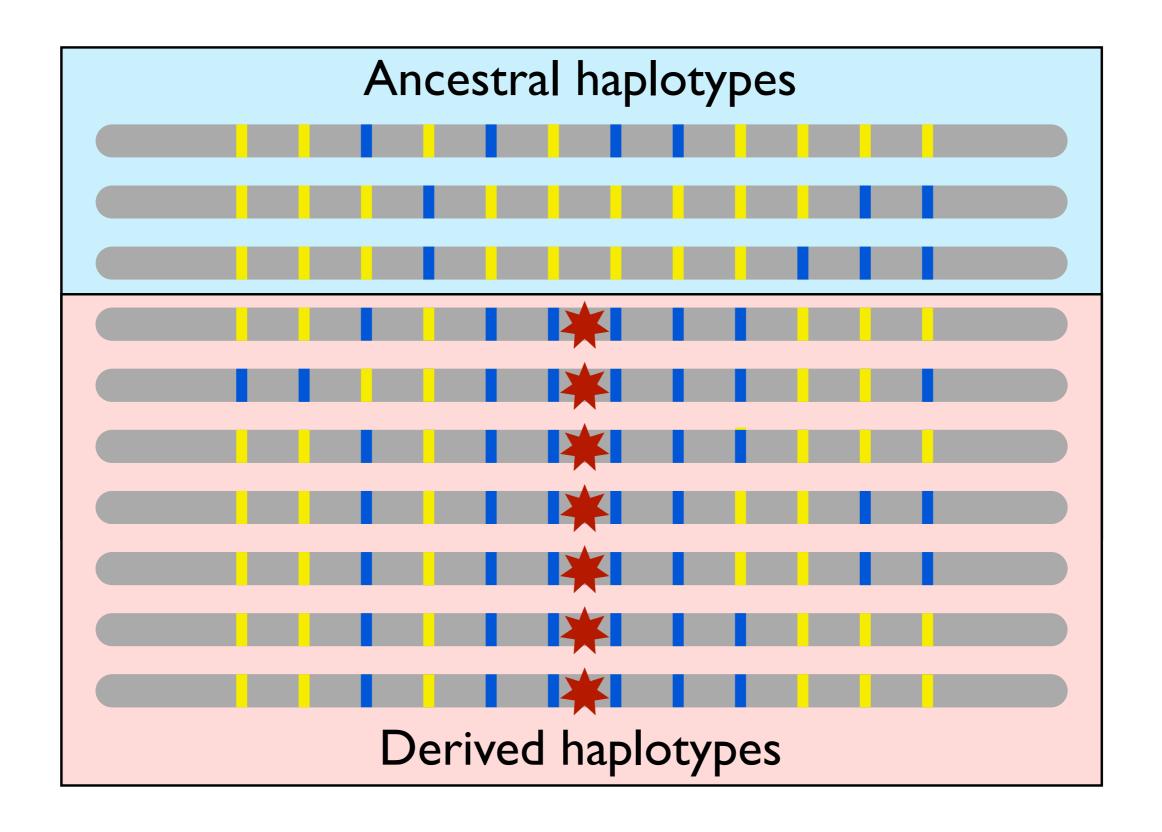
$$EHH_c(x) = \sum_{h \in \mathcal{H}(c,x)} rac{inom{n_h}{2}}{inom{n_c}{2}}$$
 # of ways to choose two h haplotypes
$$\binom{n}{2} \coloneqq 0 \ \ \forall n < 2$$

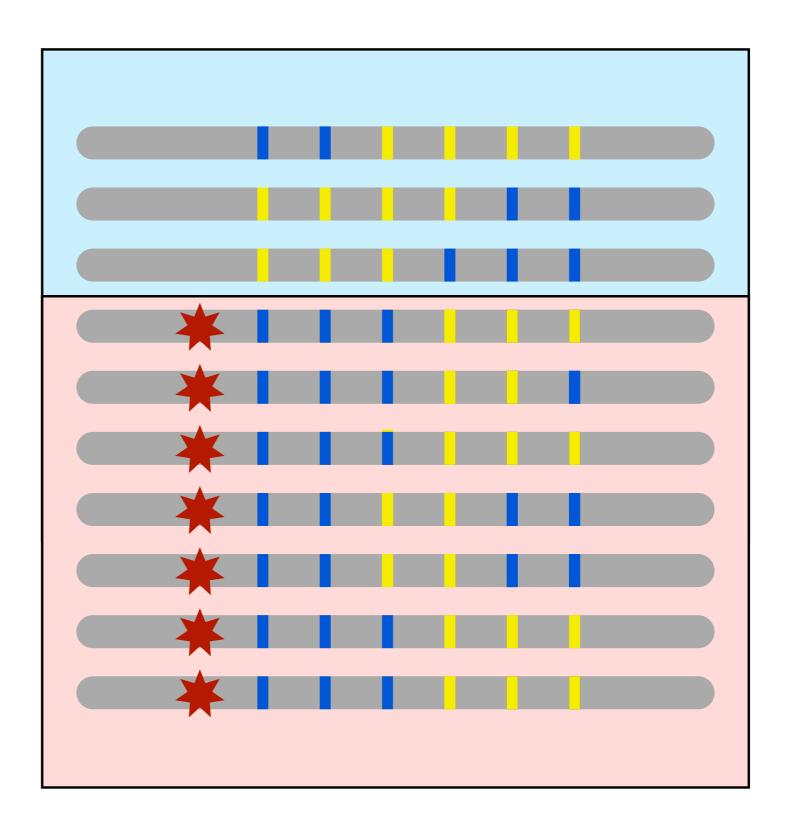
• If $EHH_c(x)$ is the extended haplotype homozygosity of the core haplotype c out to marker x, then

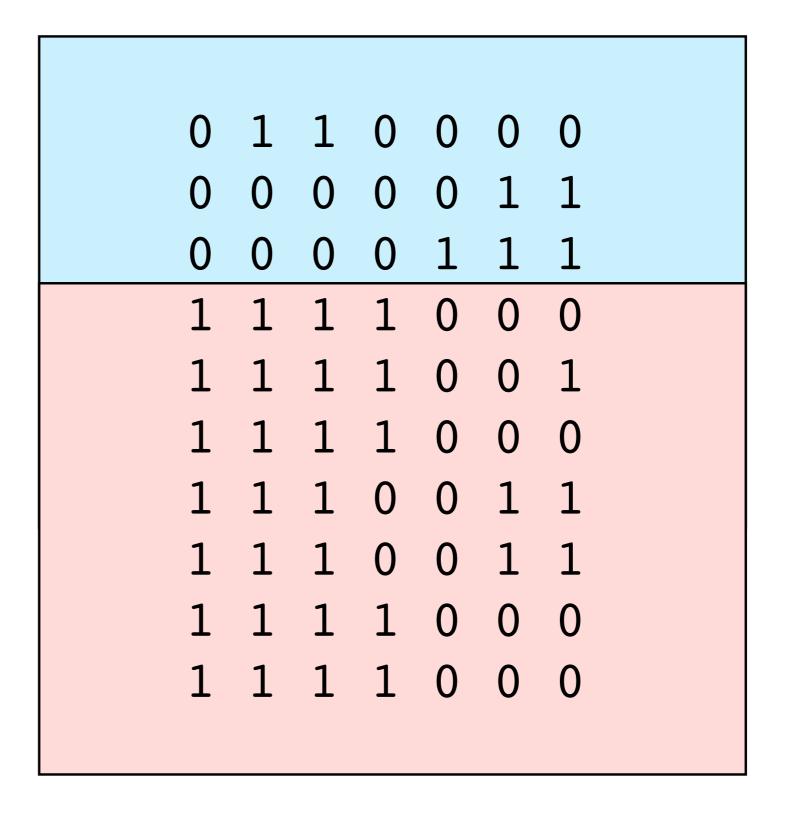
$$EHH_c(x) = \sum_{h \in \mathcal{H}(c,x)} \frac{\binom{n_h}{2}}{\binom{n_c}{2}} \overset{\text{\# of ways to choose two h haplotypes}}{} \text{\# of ways to choose two core haplotypes}$$

$$\binom{n}{2} \coloneqq 0 \ \ \forall n < 2$$

 Notice that EHH at the core haplotype is necessarily 1 and that it tends to 0 as the number of distinct haplotypes tends to infinity.





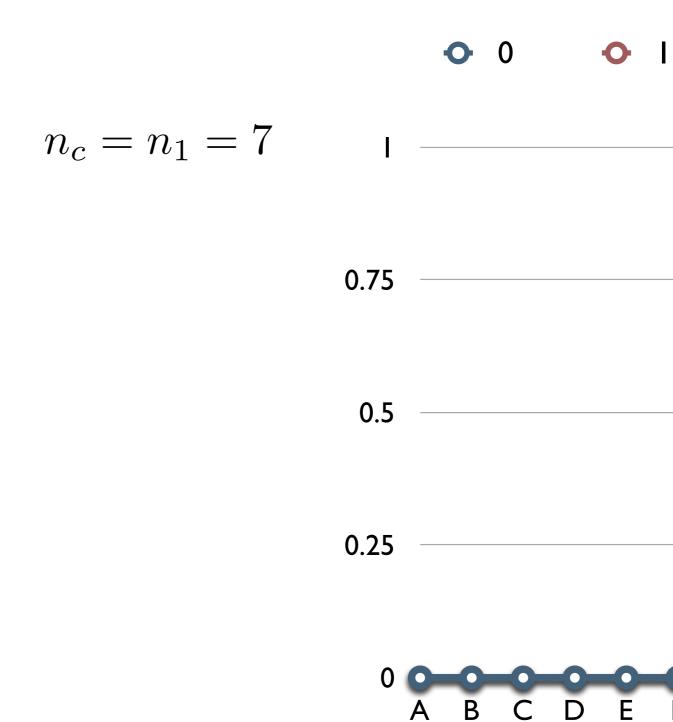


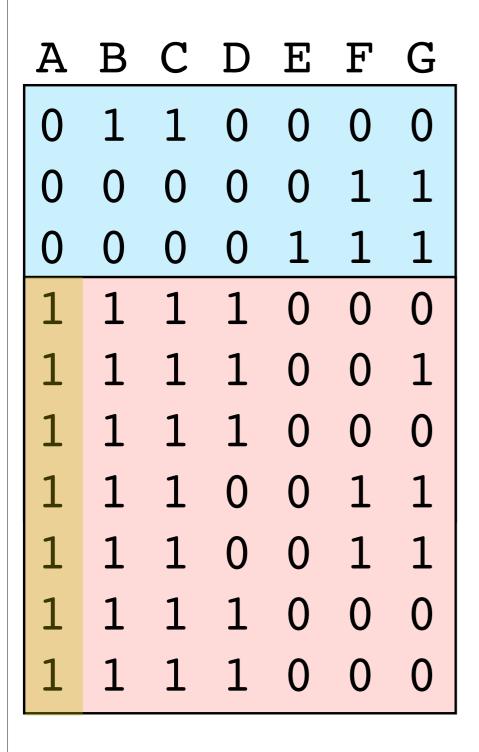
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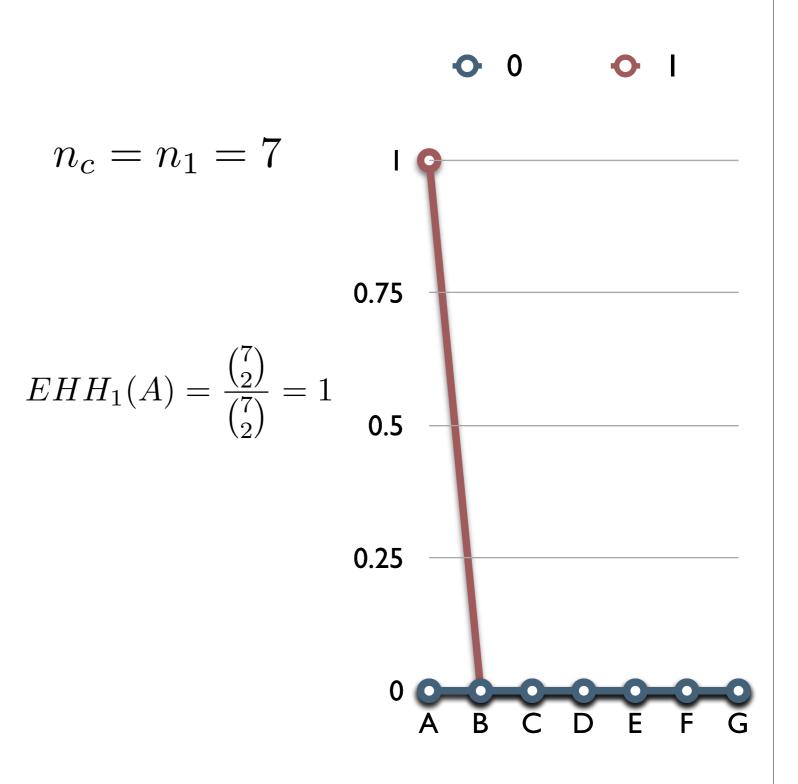
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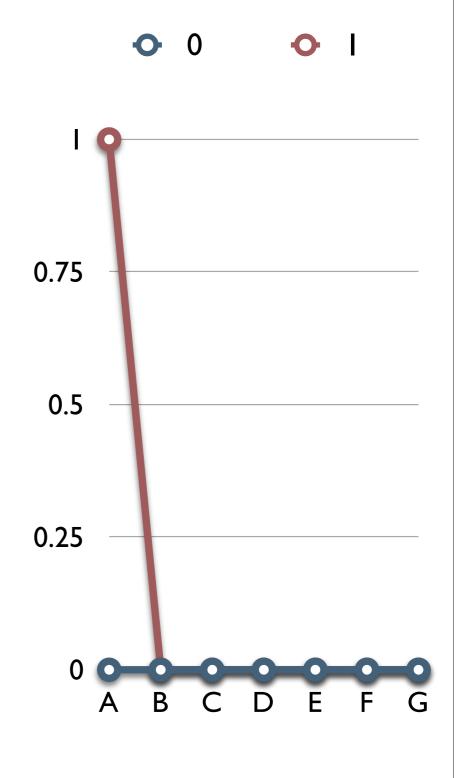
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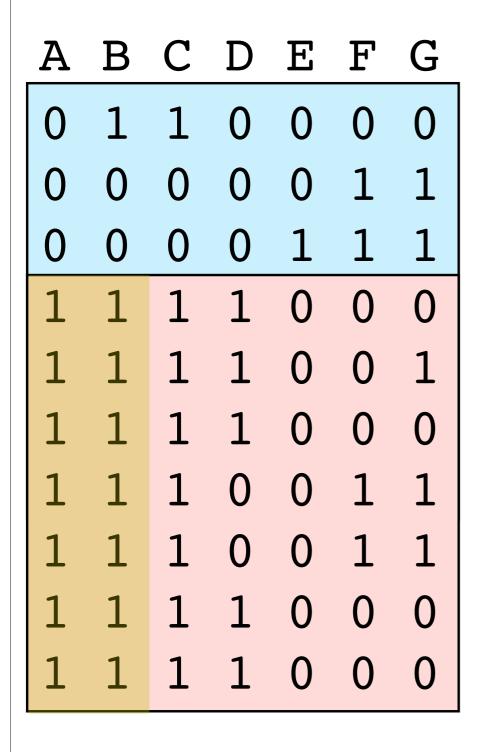


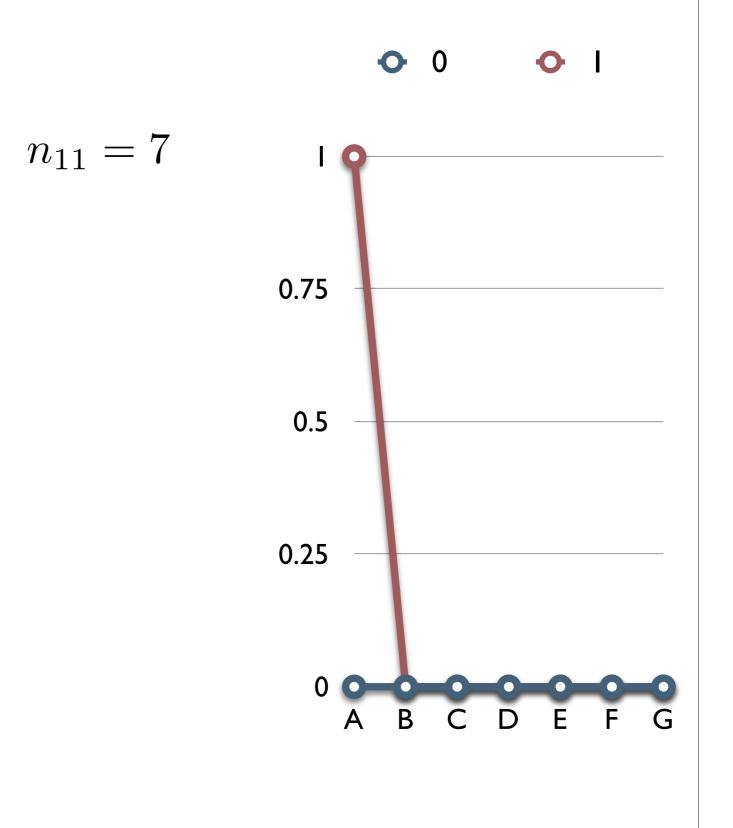


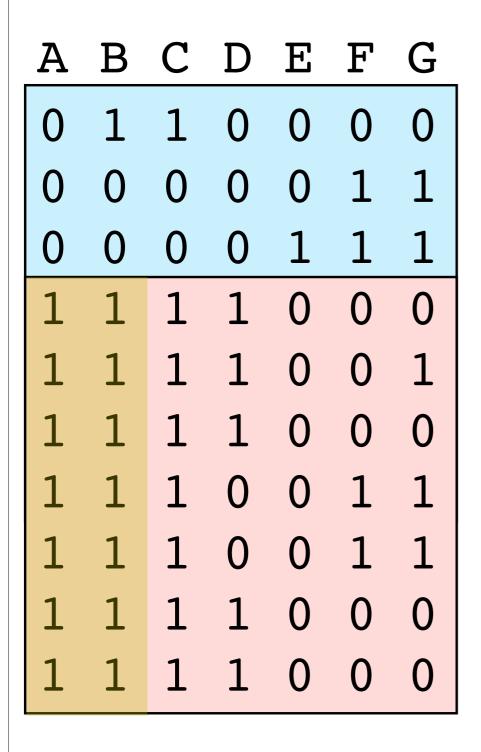


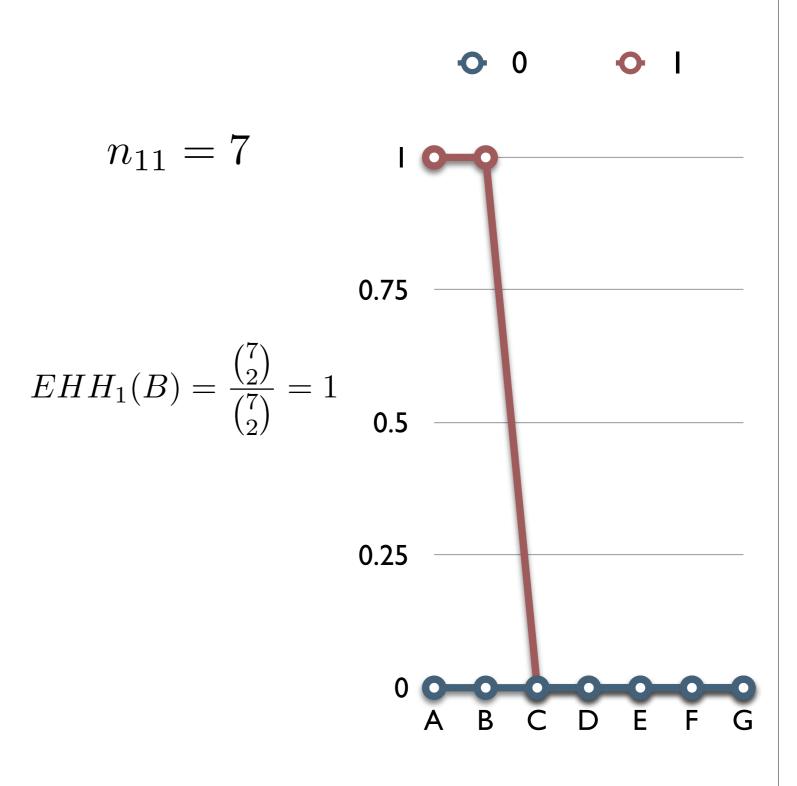
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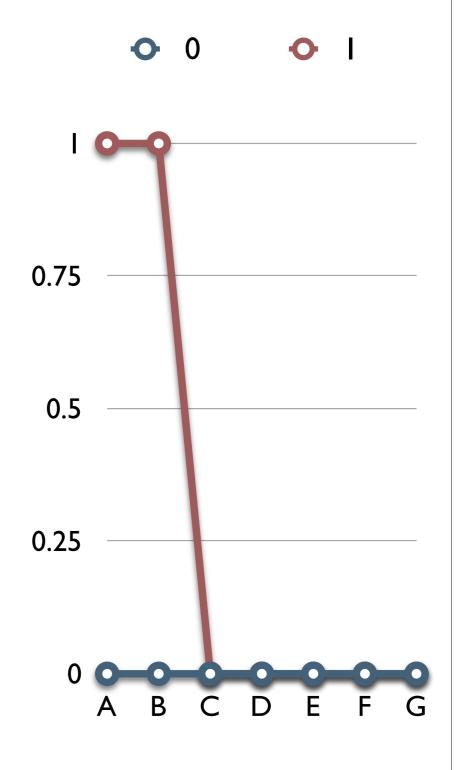


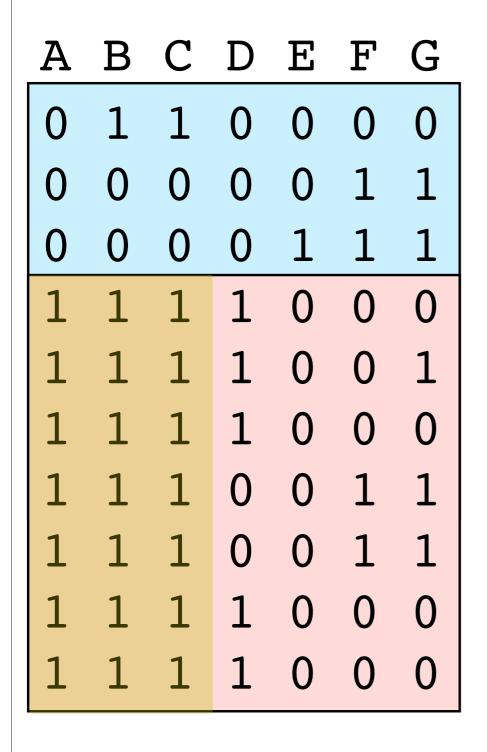


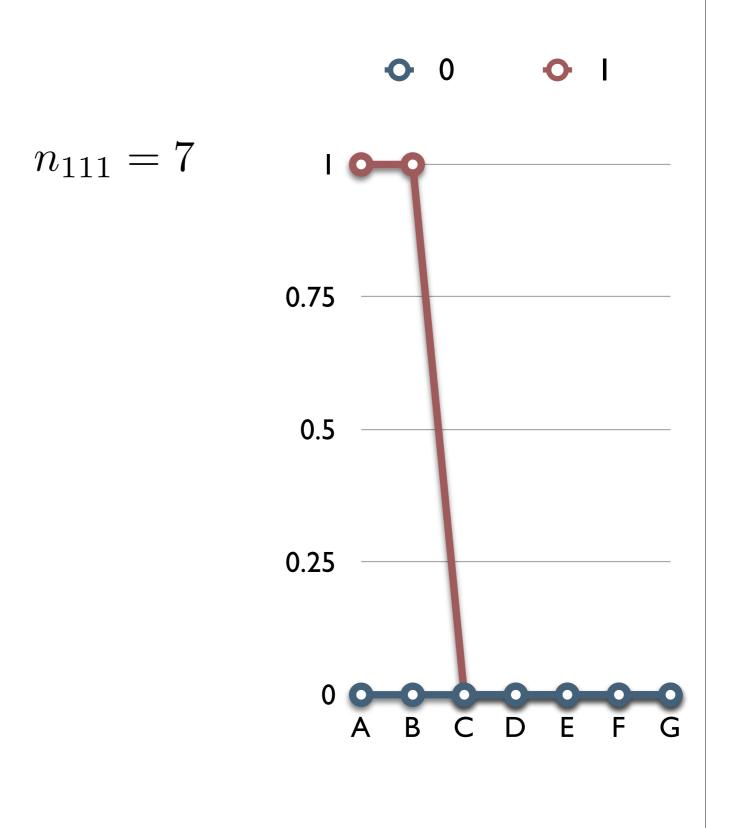


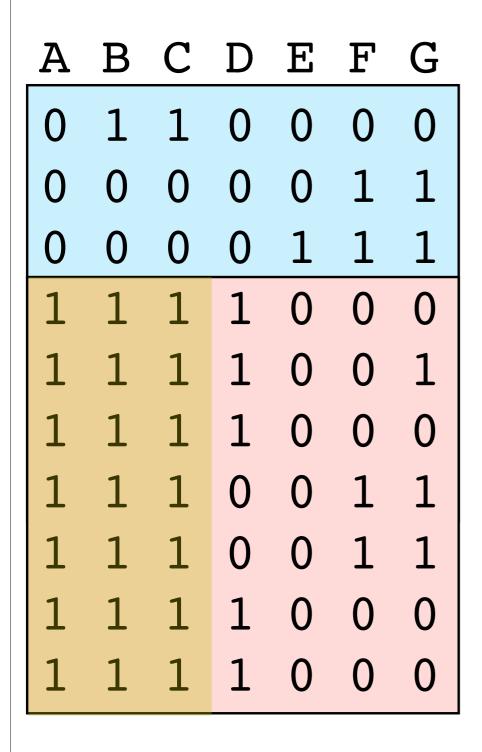


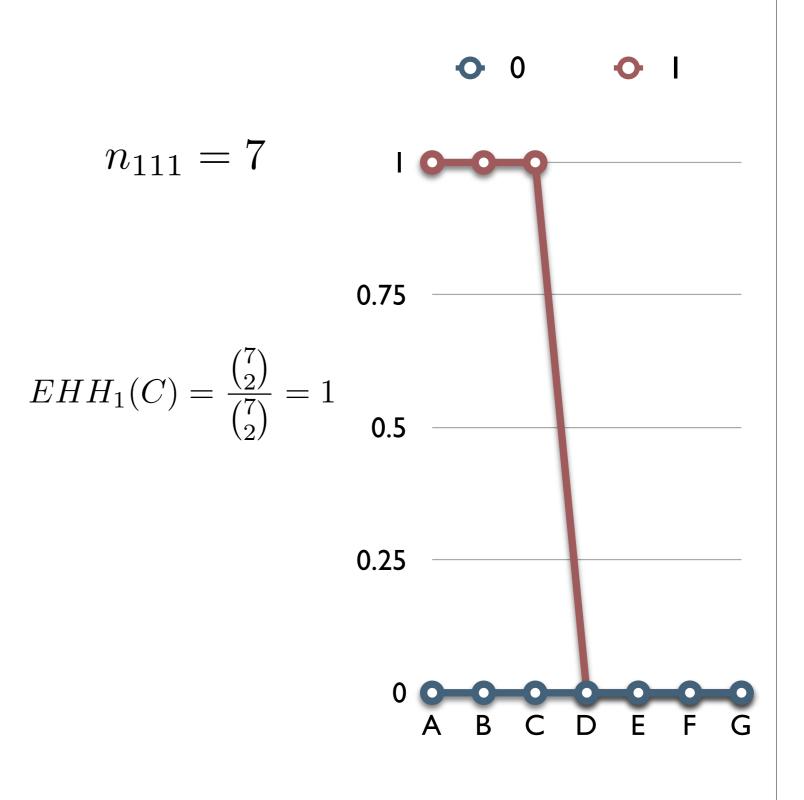
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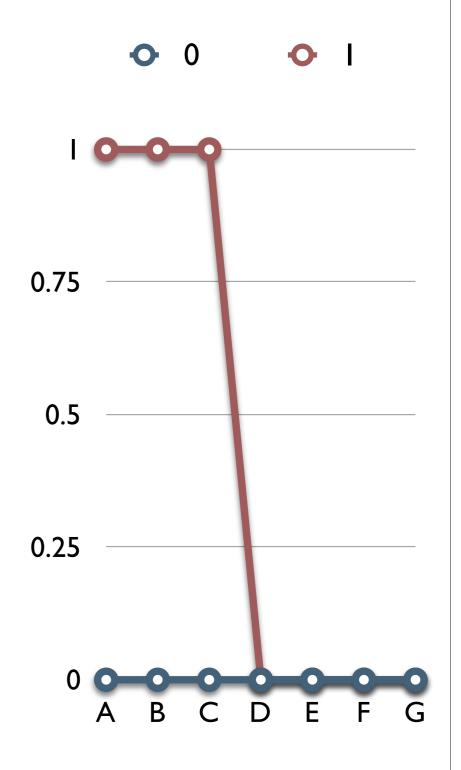




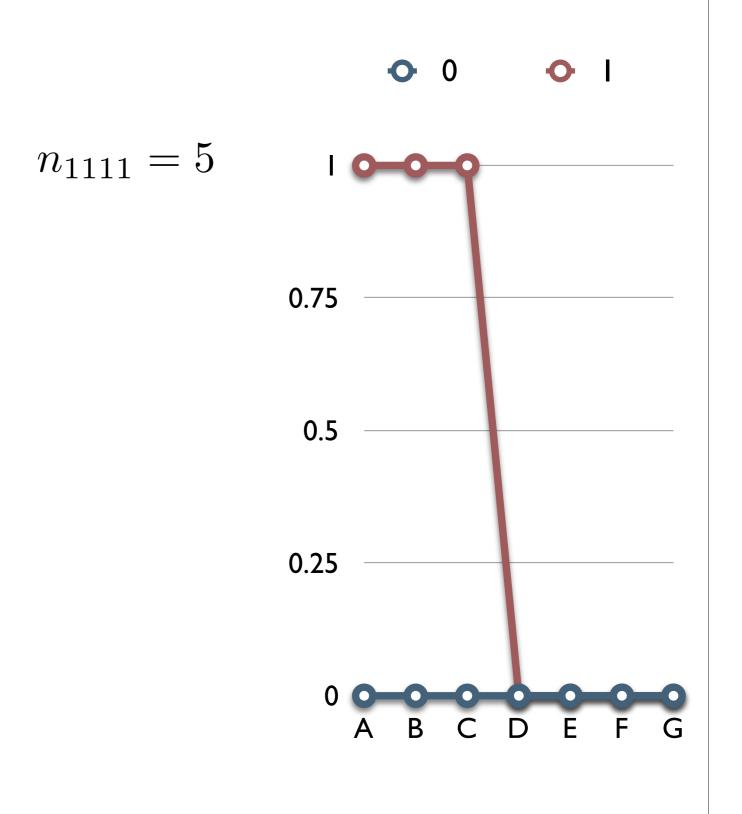




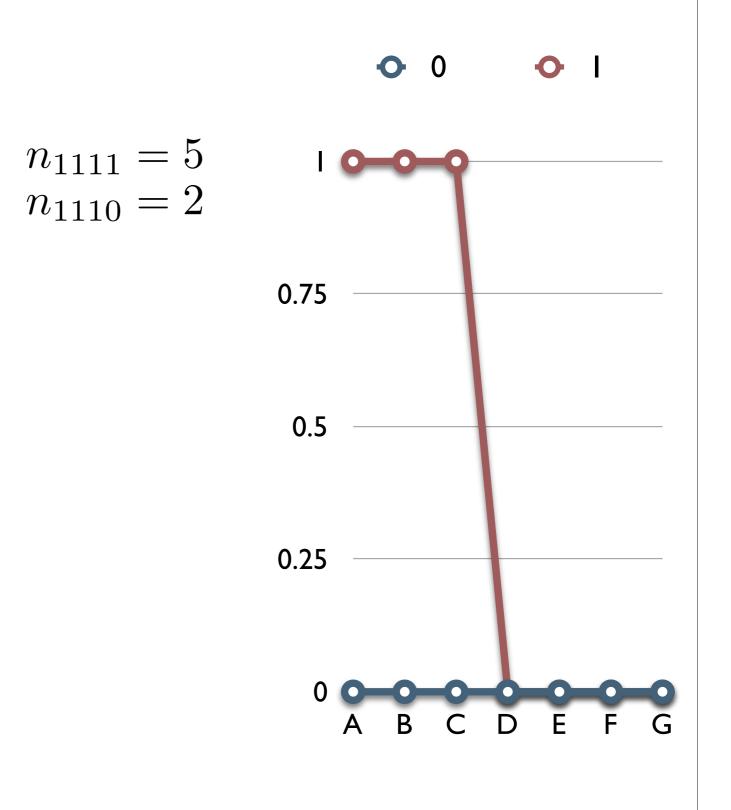
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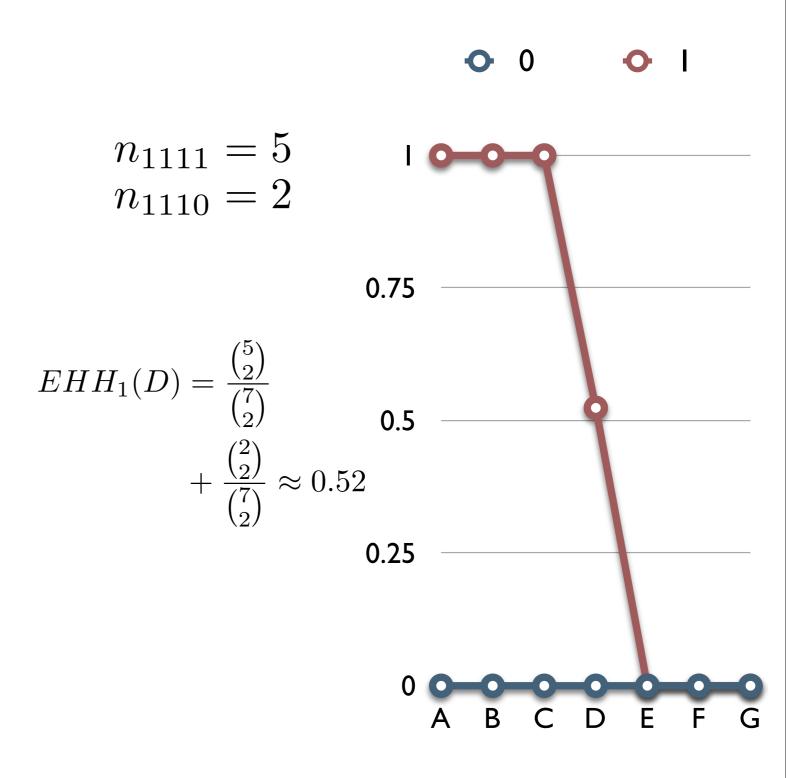
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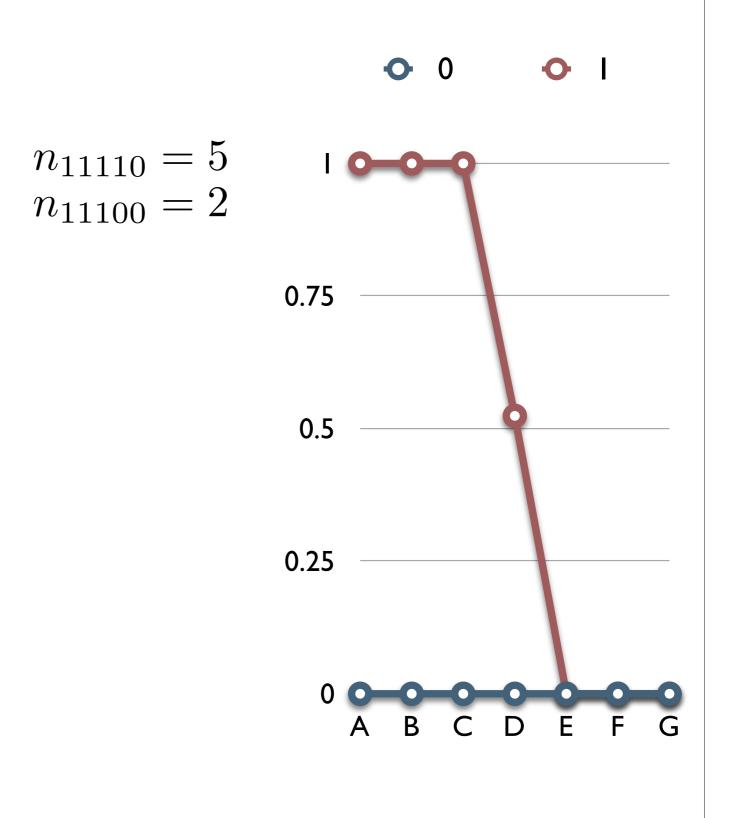
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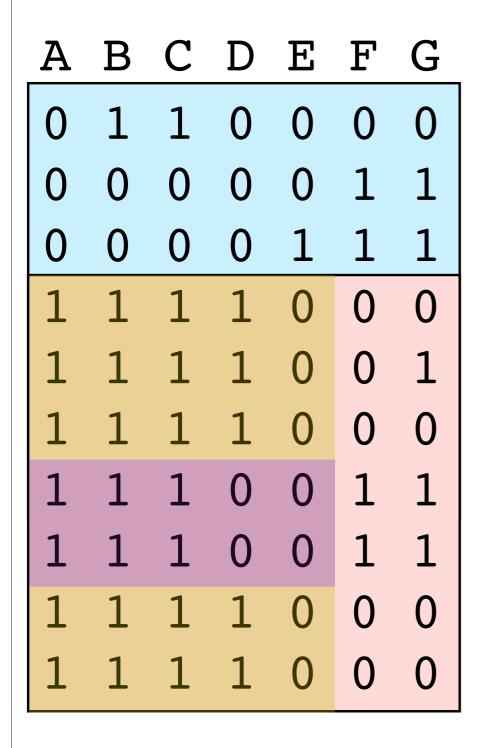


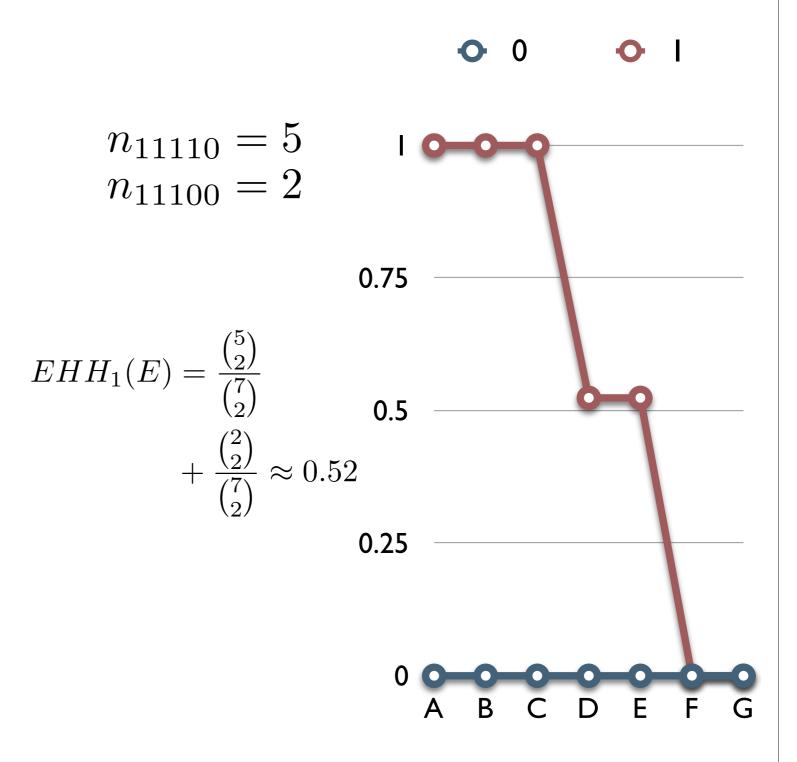
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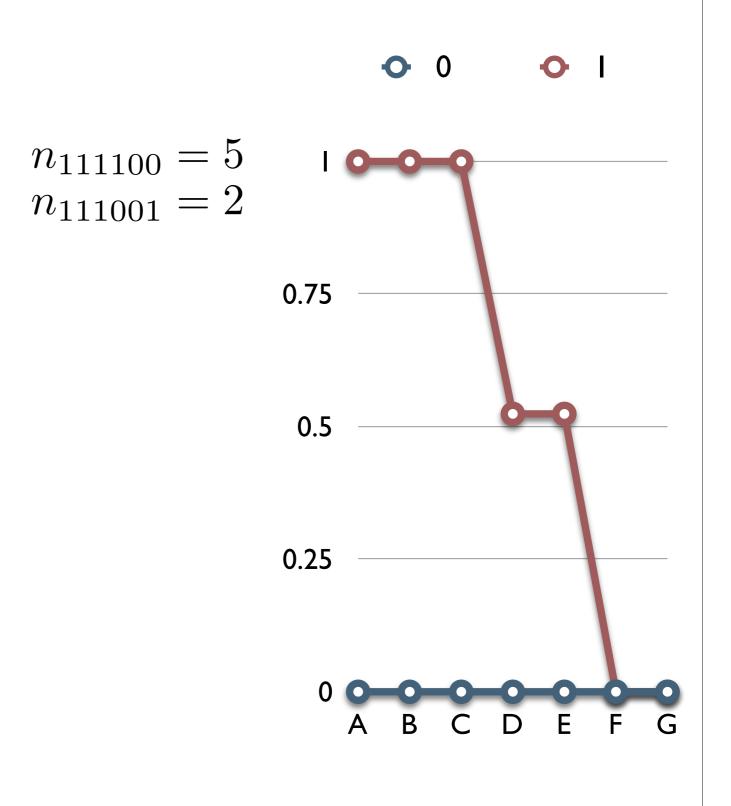
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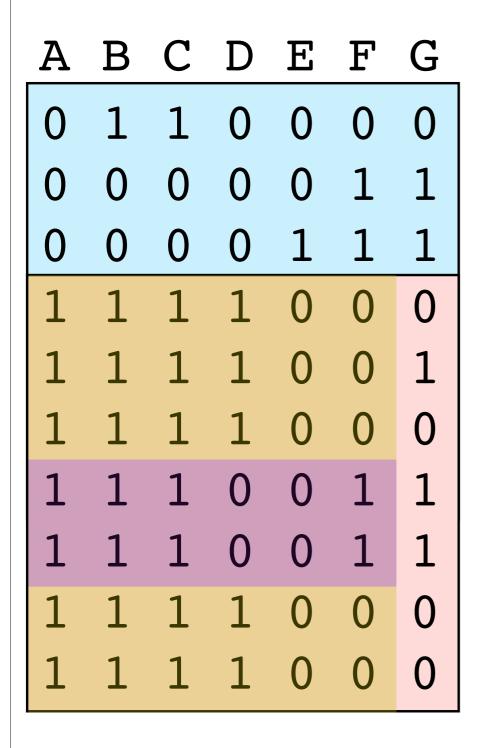


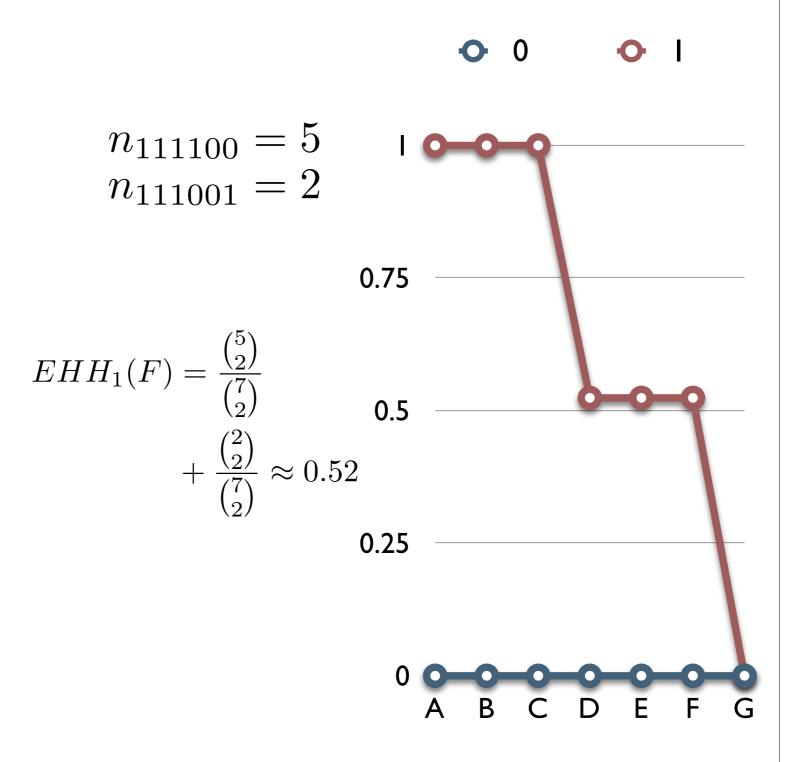


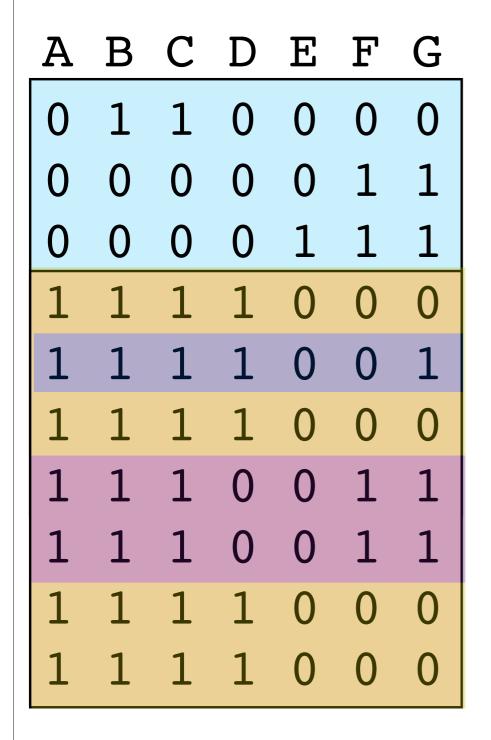


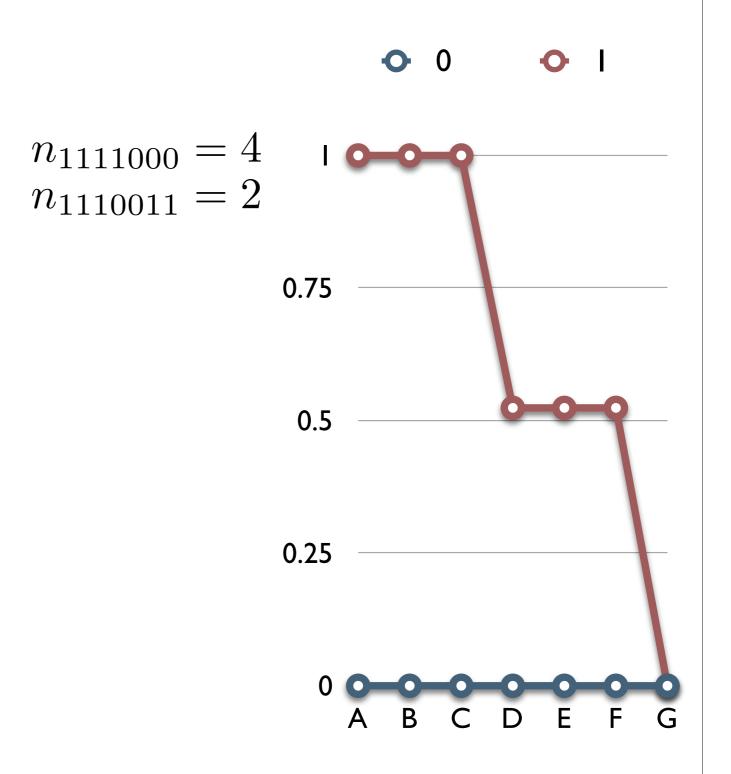
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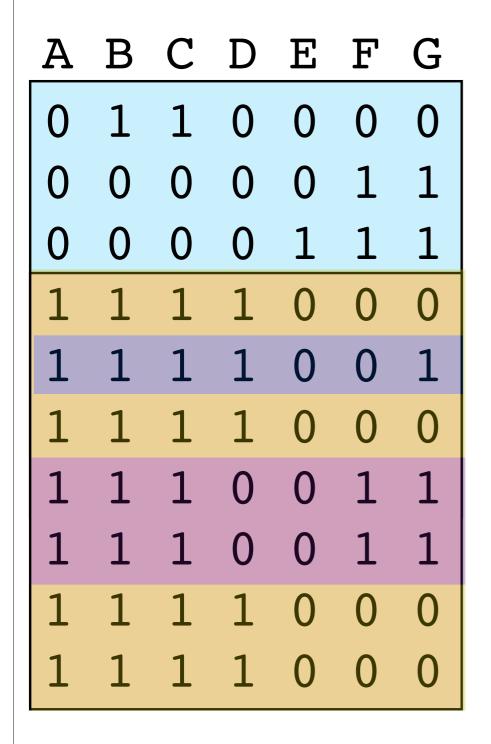


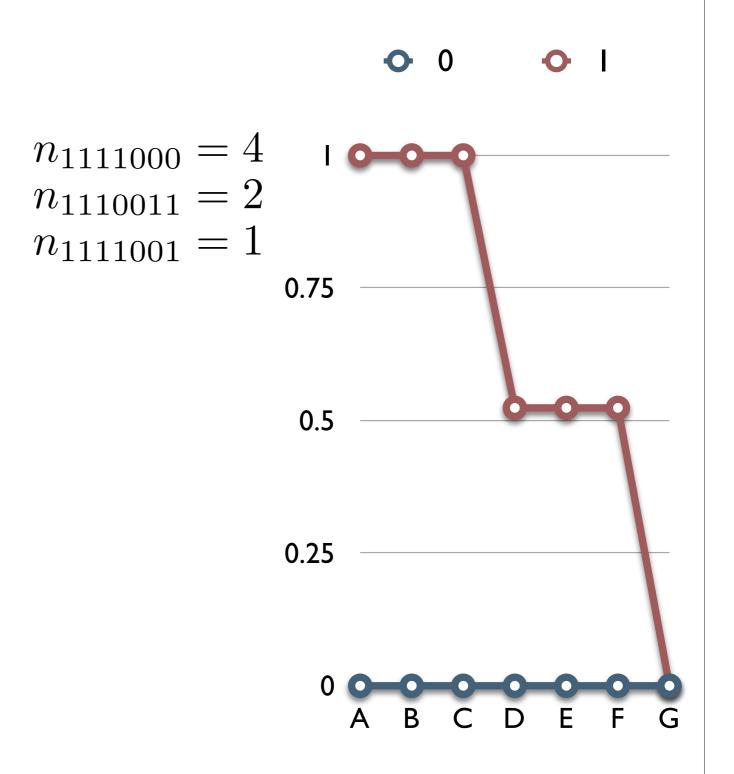


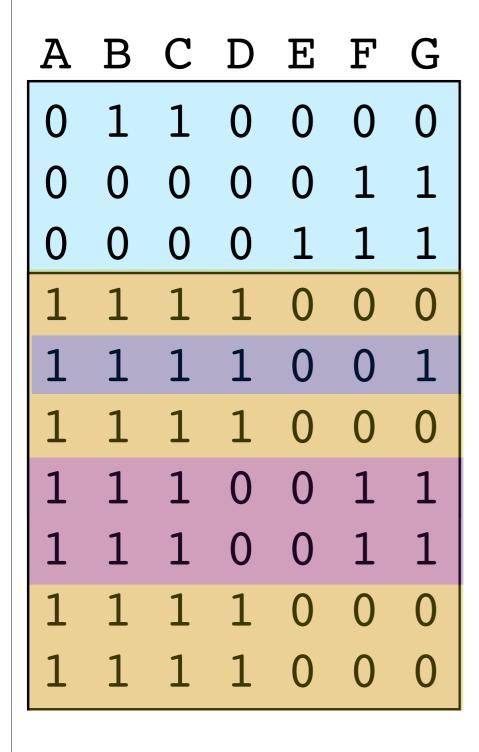


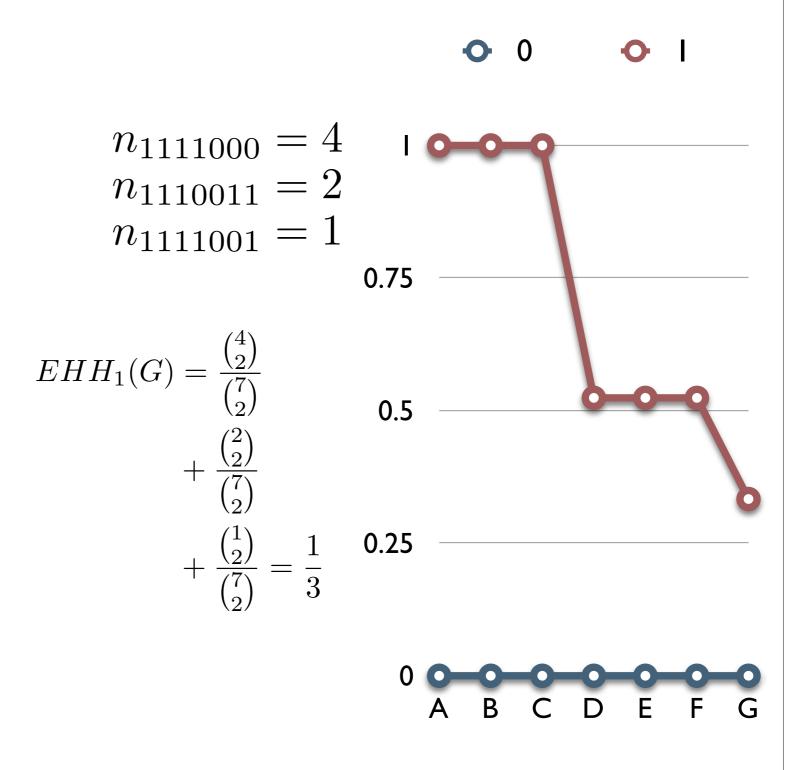




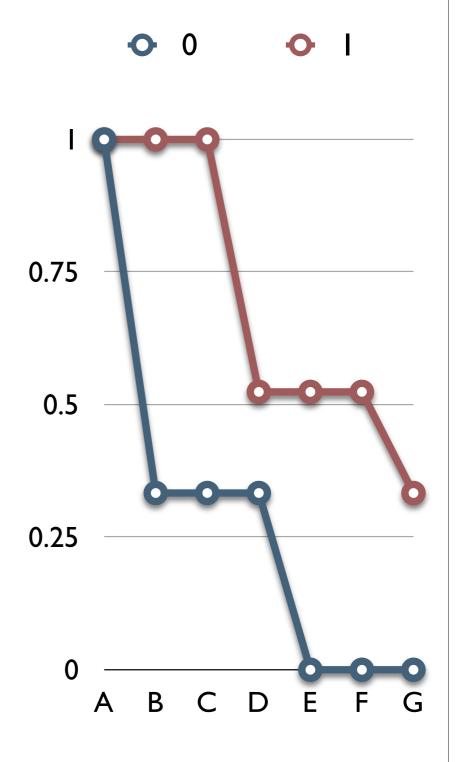


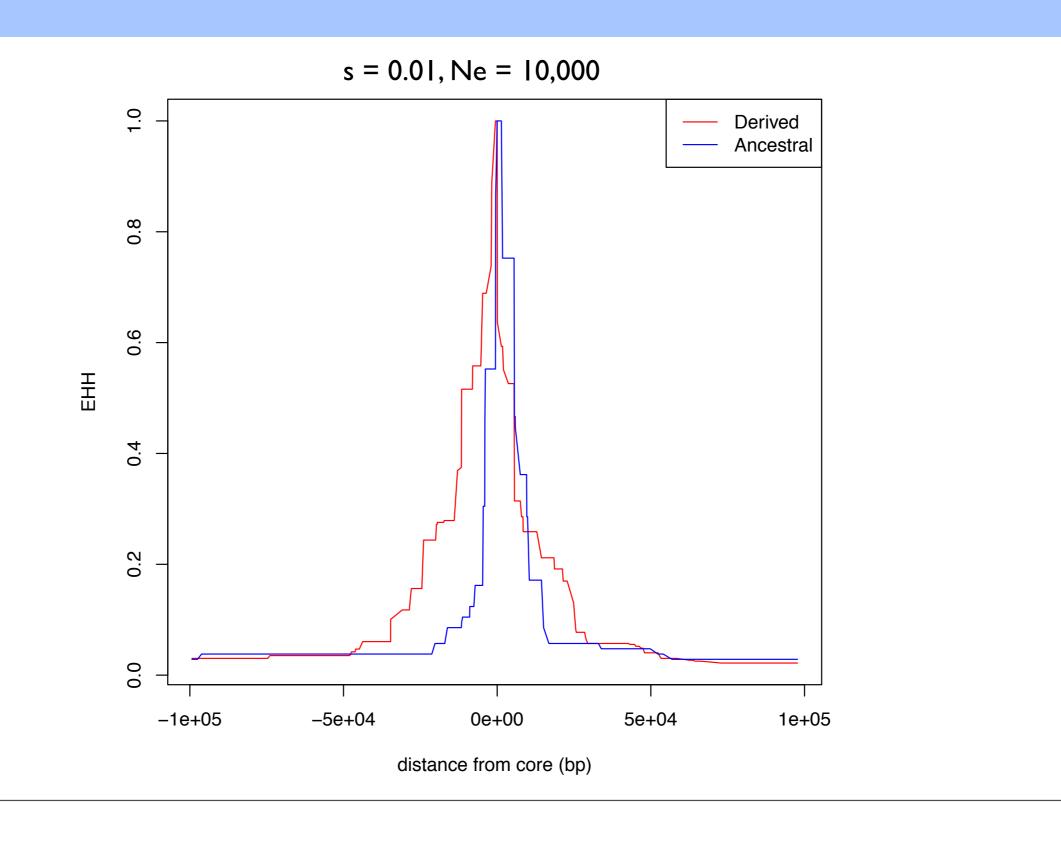


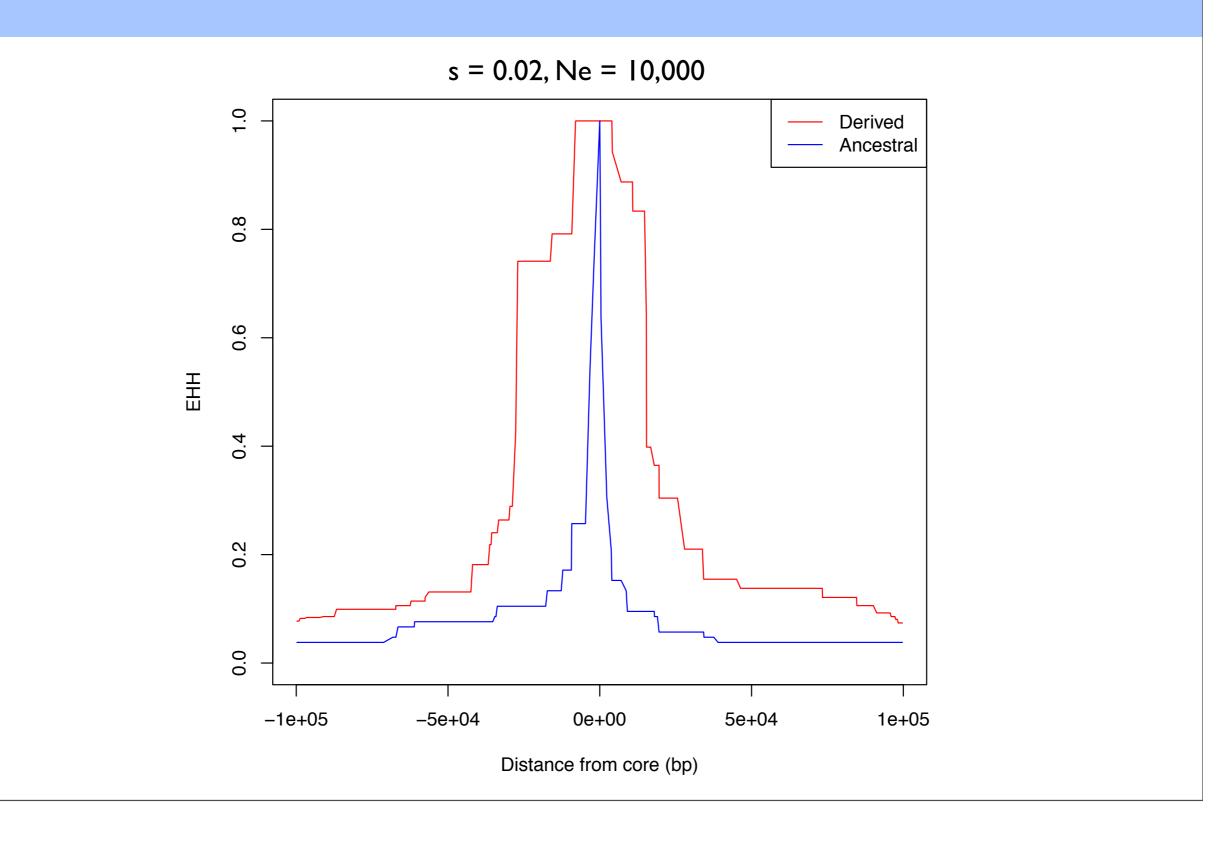


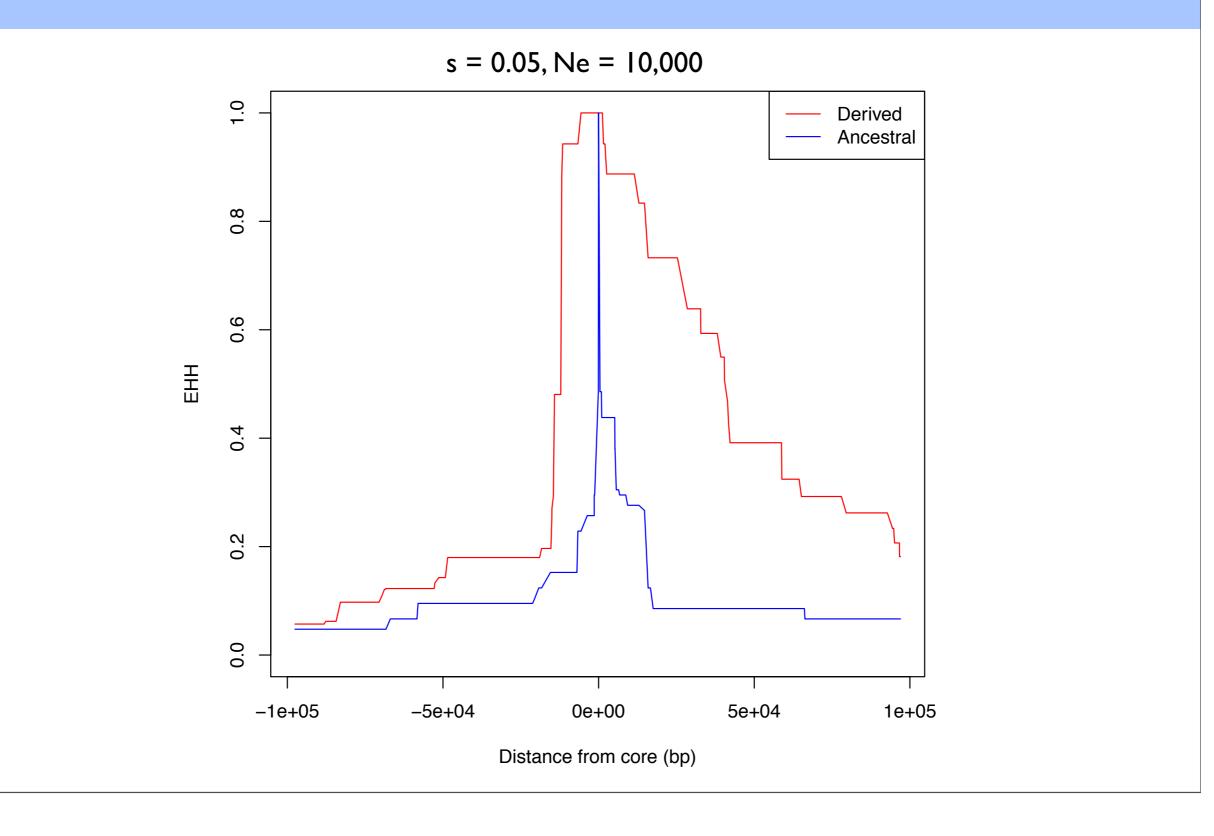


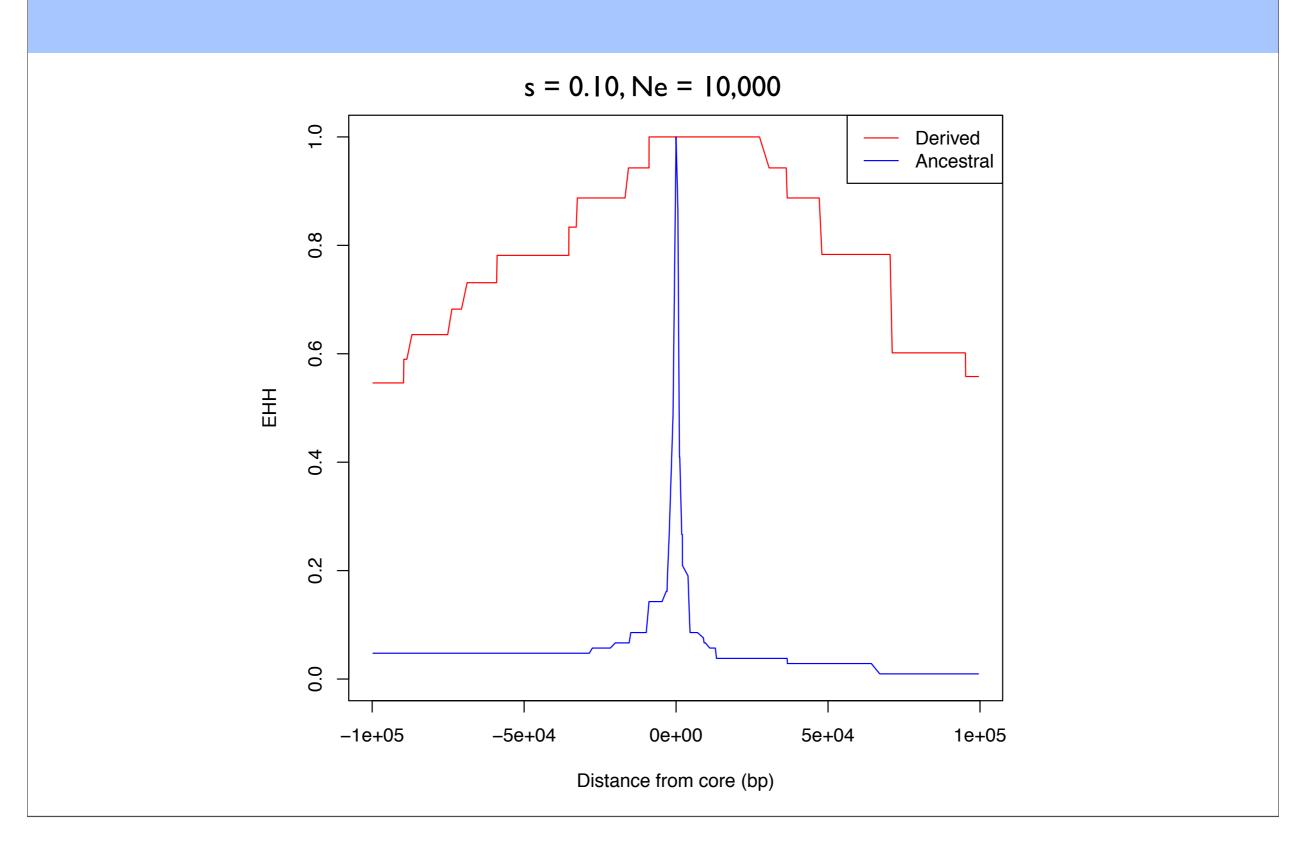
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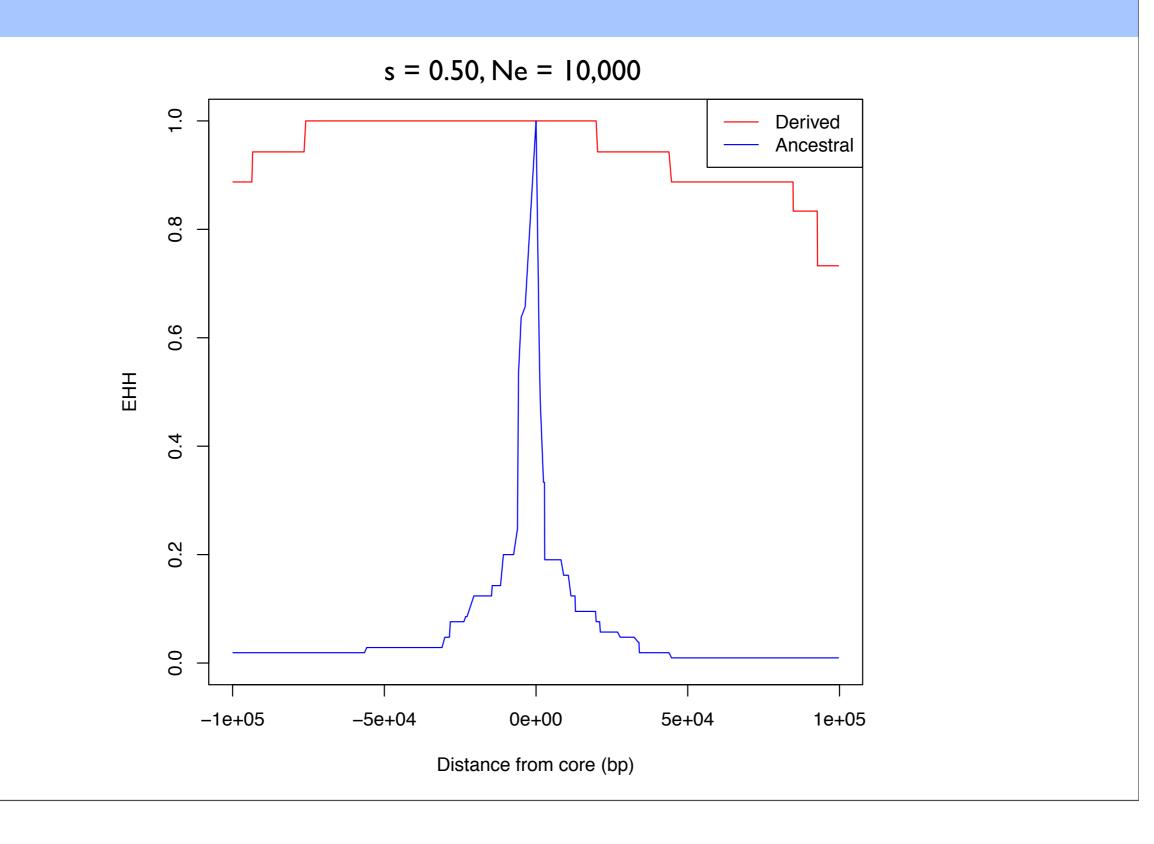












EHH

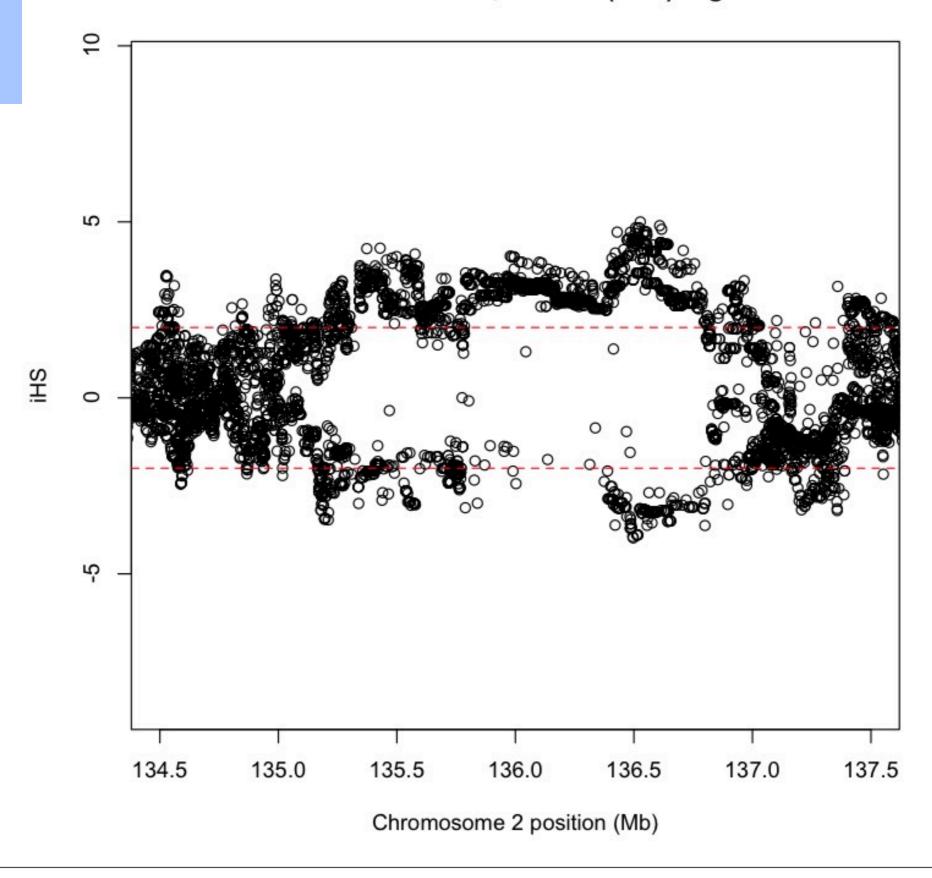
- When querying a specific region of the genome, for each core haplotype, calculate EHH for successively longer surrounding haplotypes.
- Statistical significance is determined by comparing EHH scores to neutral simulations and random control regions of the genome.

Genome-wide scans

- The EHH approach does not lend itself to a genomewide scan.
- Voight, et al. (2006) create a genome-wide scan statistic based on EHH called integrated Haplotype Score (iHS).

Voight, et al. (2006) PLoS Biology

CEU TGP Phase 3, lactase (LCT) region



Computational Tips

- Associative arrays for haplotype comparison and counting
 - O(log N)
- Instead of computing EHH until the end of the data stop after a certain distance away from the core
 - Either EHH < 0.05 or distance from core > 1Mb
- Multithreading
 - Adjacent SNPs don't rely on each other to complete calculation
 - Compute adjacent scores on separate threads

Szpiech and Hernandez (2014) Molecular Biology and Evolution

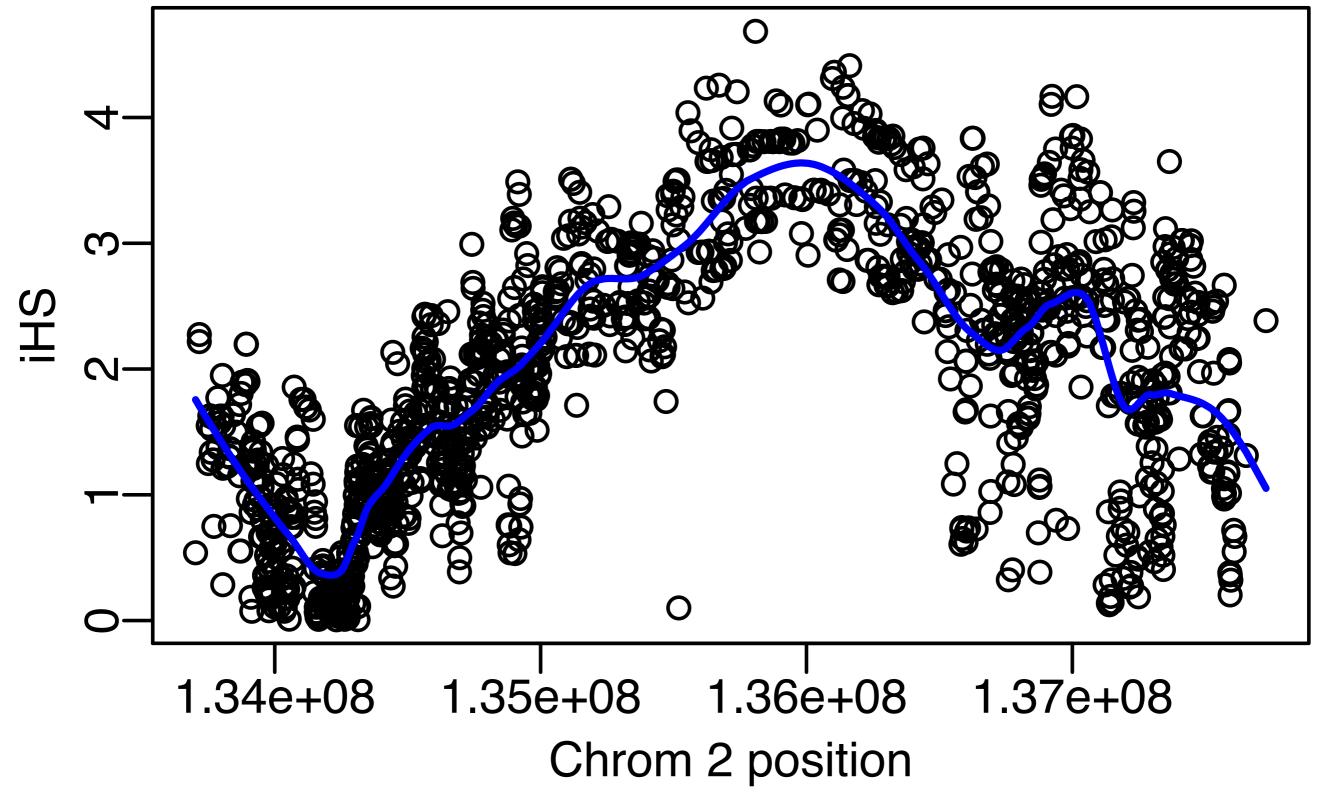
Caveats

- Power may be overstated.
 - If a large proportion of the genome is non-neutral, we lose power to detect the weakest selected variants because of genome-wide normalization.
- iHS no formal test to decide significance.
 - Take top 1% of signals
- XP-EHH more sensitive to demographics
 - i.e. comparing populations with serial bottlenecks separating them
- Important to combine multiple lines of evidence!

Breakout Groups!! Running iHS with selscan

- Open up your command prompt (i.e., rev your engines)
- Let's give iHS a go!
- Let's consider the LCT gene.
- Follow commands in Day3_AM2_CMD.txt
 - You will need selscan.zip
 - In terminal run:
 - selscan ...
 - norm ...
 - Plot it in R!

iHS



Other populations??

- Now run selscan on the YRI population
- YRI is a sample of individuals from Yoruba, Nigeria, where they
 do not have a long tradition of domesticating cows.
- Update the selscan commands by replacing "CEU" with "YRI"
- Breakout Groups!!

Do you think there is selection in this region in the YRI population?

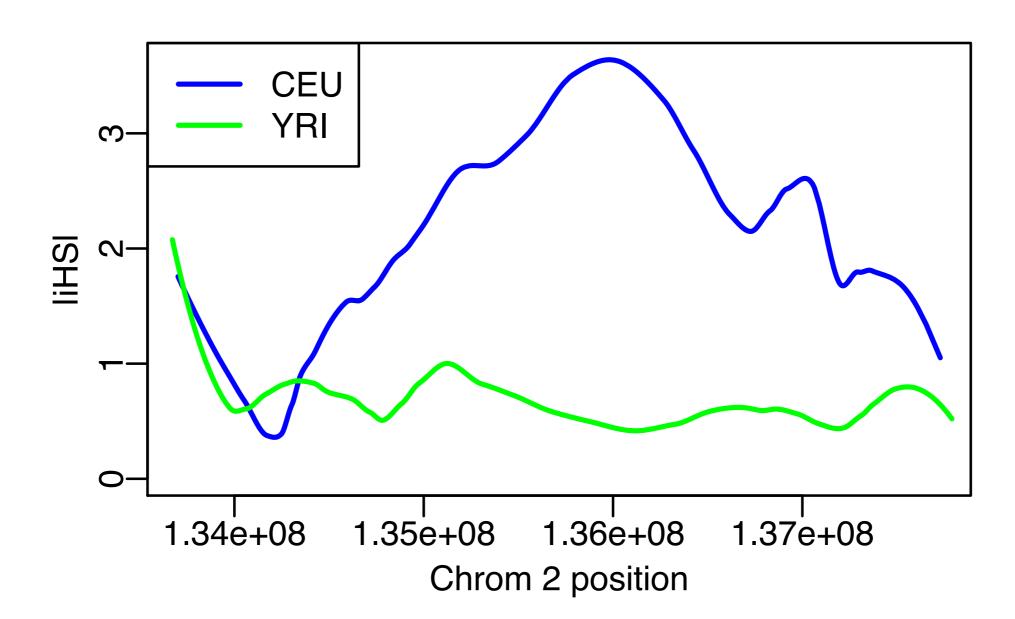
Yes!

No!

Unclear

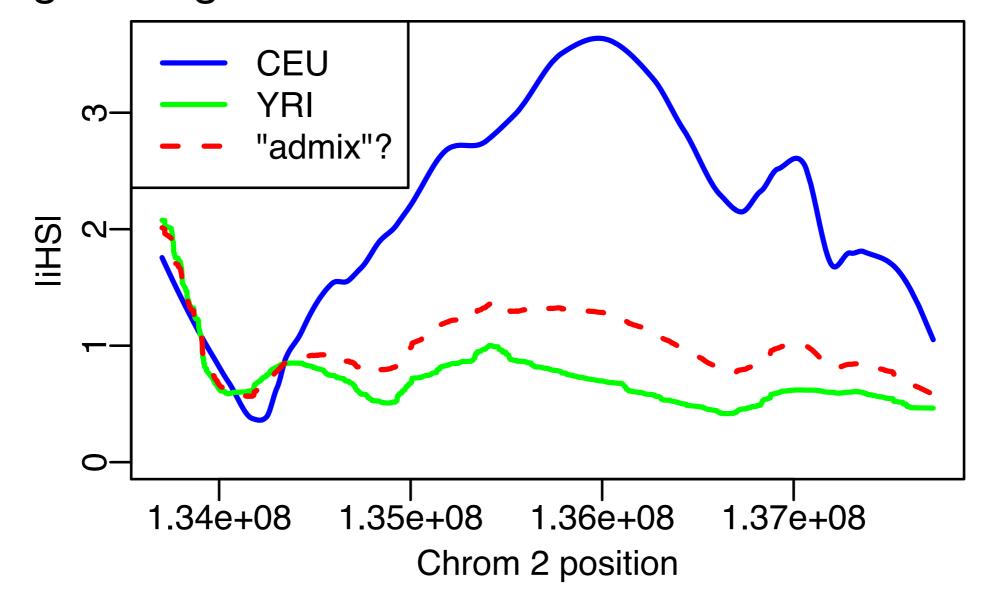
Other populations??

• "CEU" vs "YRI"



What about admixture?

- African American genomes contain admixture with African ancestry (~80%) and European ancestry (~20%).
- ASW is one sample of African Americans (from the Southwest)
- One guess might be that it should be intermediate

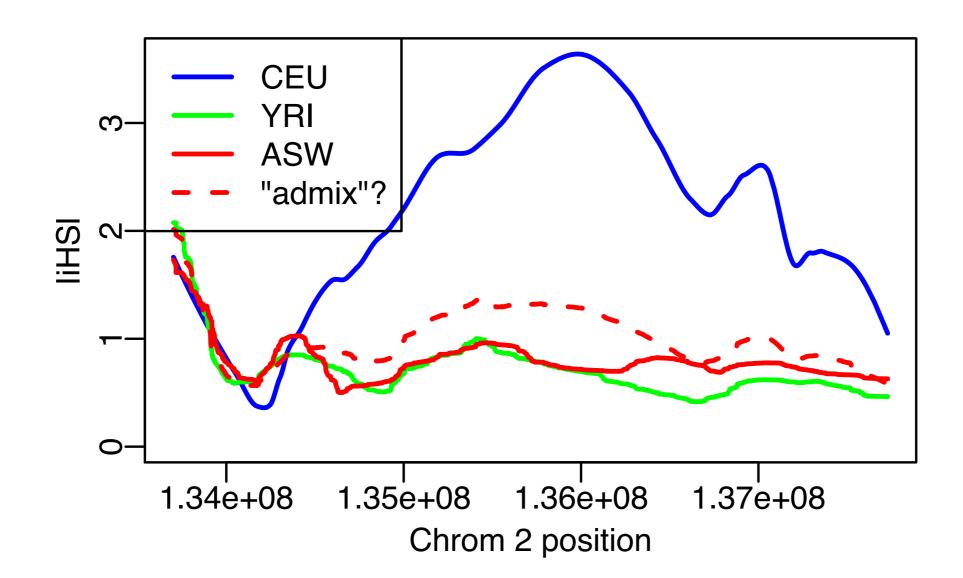


Other populations??

- Now run selscan on the ASW population
- Update the selscan command by replacing "CEU" with "ASW"
- Breakout groups!!

Other populations??

- Now run selscan on the ASW population
- Update the selscan command by replacing "CEU" with "ASW"
- In these data, ASW is much more similar to YRI than "expected".



Summary

- iHS is one example of a statistic geared toward detecting a "classic sweep".
- It is based on the idea that a new mutation has been selected, and quickly spread through the population.
- selscan is one piece of software that can run many different selection statistics in an efficient manner.

- Open your terminal/command prompt!
- Change to the new selscan directory
- For example:
 - cd ~/Desktop/selscan/
- There should 4 subdirectories:
 - rhernandez\$ lsdata linux osx win
- Change Directory to where the data are:
 - cd data

- All the commands we are running can be found in the selscan_CMD.txt file.
- Copy the appropriate executable to the data directory:
- osx:
 - cp ../osx/selscan .
- linux:
 - cp ../linux/selscan .
- Windows:
 - copy ..\win\selscan.exe .

• Test that it works:

```
osx/linux: ./selscan (Win: selscan.exe)
selscan v1.1.0b
ERROR: Must specify one and only one of
EHH (-ehh)
iHS (--ihs)
XP-EHH (--xpehh)
PI (--pi)
nSL (--nsl)
```

- iHS requires 2 files, a map file and a hap file.
 - --map <string>: A mapfile with one row per variant site.
 - Formatted with 4 columns:
 - <chr#> <locusID> <genetic pos>
 <physical pos>
 - --hap <string>: A hapfile with one row per haplotype, and one column per variant.
 Variants should be coded 0/1.

- Now run it!
 - All in one line type:

```
• ./selscan (Win: selscan.exe)
                  --ihs
                  --map CEU.chr2.map
                  --hap CEU.chr2.ihshap
                  --out CEU.chr2
selscan v1.1.0b
Opening ../data/CEU.chr2.hap...
Loading 224 haplotypes and 1971 loci...
Opening ../data/CEU.chr2.map...
Loading map data for 1971 loci
--skip-low-freq set. Removing all variants < 0.05.
Removed 359 low frequency variants.
Starting iHS calculations with alt flag not set.
              ===========================>
```

Normalize

- All in one line type:
 - ./norm
 - --ihs
 - --files CEU.chr2.ihs.out bg.ihs.out

```
norm v1.1.0aYou have provided 2 output files for joint normalization.
```

Opened ../data/CEU.chr2.ihs.out

Opened ../data/bg.ihs.out

Total loci: 666285

Reading all frequency and iHS data.

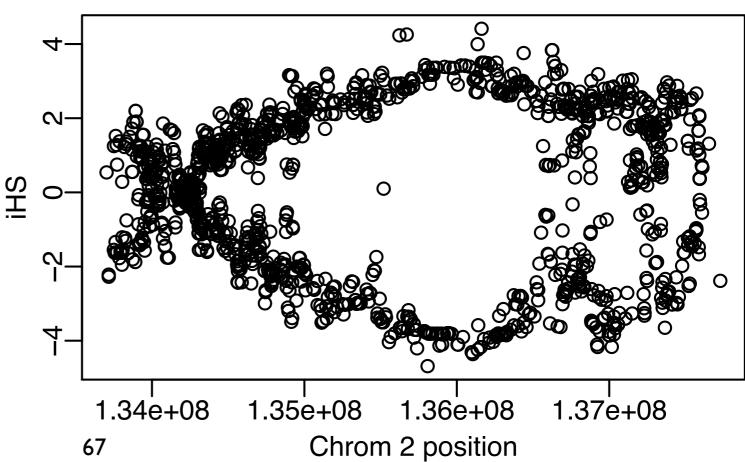
Calculating mean and variance per frequency bin:

iHS

- Now let's plot it!
- Open R.
- Read in data for CEU:

```
setwd("cd ~/Desktop/selscan/data")
CEU=read.table("CEU.chr2.ihs.out.100bins.norm")
```

plot(CEU[,2], CEU[,7])



iHS

- Often analyze absolute value, and smooth it out.
- My preferred method for smoothing is using loess

```
SP=0.2 #this is the span, a parameter you can change (higher = more smoothing)

CEU.x=CEU[,2]; #the x-coordinates in Mb

y=abs(CEU[,7]) #iHS is actually the absolute value

CEU.loess=loess(y~CEU.x,span=SP,data.frame(x=CEU.x,y=y)); #step 1

CEU.predict=predict(CEU.loess,data.frame(x=CEU.x)); #step 2

plot(CEU[,2], abs(CEU[,7]))

lines(CEU.x, CEU.predict, lwd=2, col='blue')
```