Pathway/ Gene Set Analysis in Genome-Wide Association Studies

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Goals

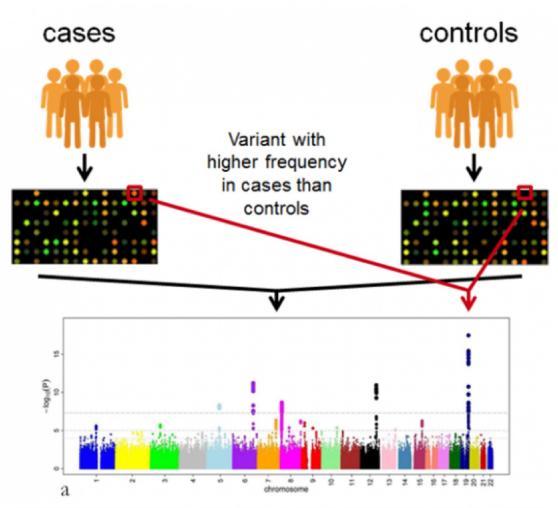
 Methods for GWAS with SNP or whole genome sequencing (WGS) chips

Many Shared Issues

 Most issues/choices/approaches discussed for microarray data are true across all "-omics"

 A few biological and technological issues that may make just "off the shelf" use of expression pathway analysis tools inappropriate

Genome-Wide Association Studies



https://www.ebi.ac.uk/training-beta/online/courses/gwas-catalogue-exploring-snp-trait-associations/what-is-gwas-catalog/what-are-genome-wide-association-studies-gwas/

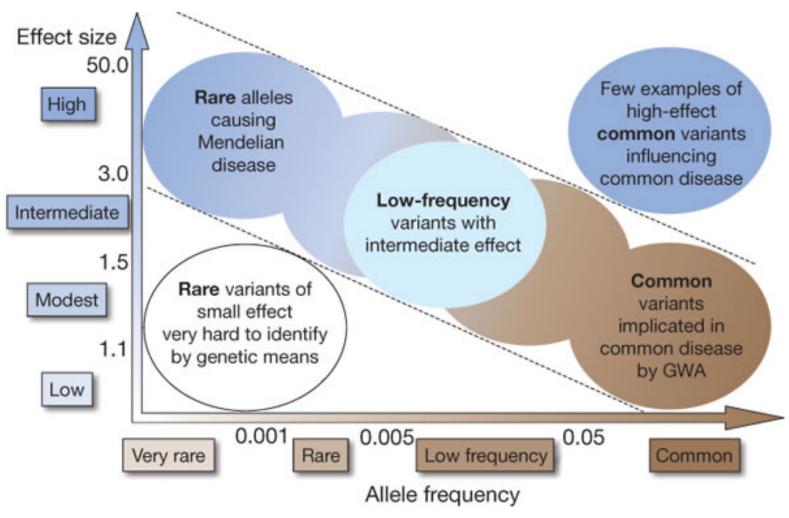
Advantages

- relatively unbiased, covers most of genome
- current cost is reasonable
- Fine mapping compared to linkage

Concerns

- missing heritability
 - Single SNPs explain little variation
- underlying assumptions not always true
 - > CDCV.....
- Standard analysis looks variant-by-variant

Feasibility of identifying genetic variants by risk allele frequency and strength of genetic effect (odds ratio).

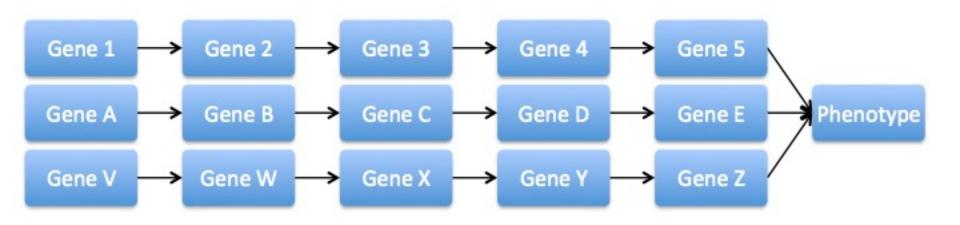


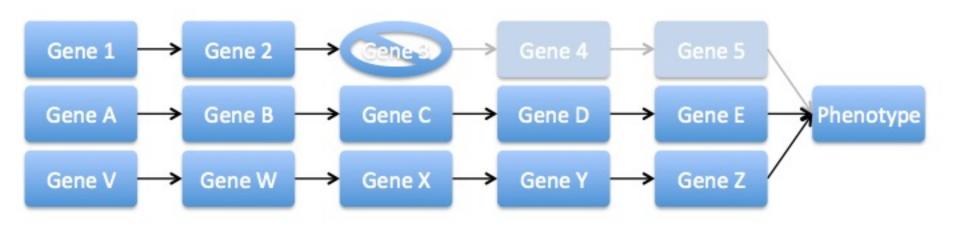
TA Manolio et al. Nature 461, 747-753 (2009) doi:10.1038/nature08494

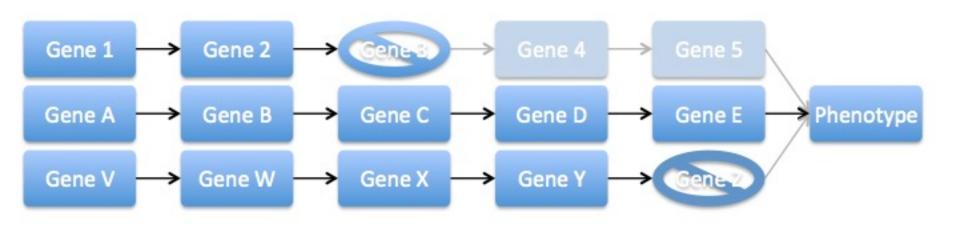
Possible Association Models

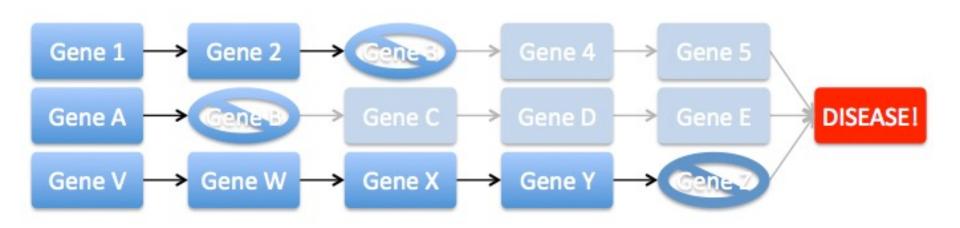
- Each of several genes may have a variant that confers increased risk of disease independent of other genes
- Several genes in contribute additively to the malfunction of the pathway

 There are several distinct combinations of gene variants that increase relative risk but only modest increases in risk for any single variant

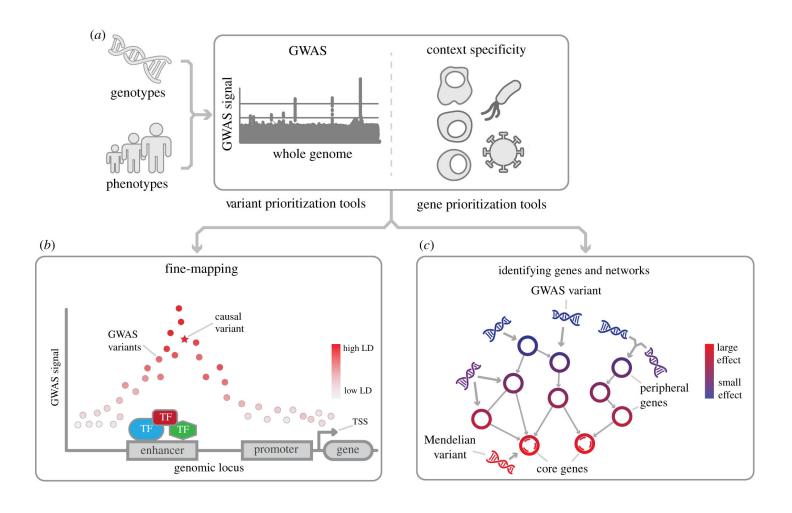








Post GWAS Era Workflow



Enrichment Testing in GWAS

- Testing pathway enrichment is possible in GWAS data
 - Many of the same issues that exist in gene expression enrichment testing occur in GWAS enrichment testing (e.g. choice of statistics, competitive vs self-contained)
- Primary difference:
 - In expression data the unit of testing is a gene
 - In GWAS data the unit of testing is a SNP
- Challenges:
 - Identifying the SNP (set) -> Gene mapping
 - Summarizing across individual SNP statistics to compute a per-gene measure
 - Correcting for LD, especially across ancestral populations
 - Correcting for gene size, pathway size, and number of variants

Candidate Gene vs. Agnostic

- Choices dependent on study goals
 - > scientific question and available data
- Candidate pathway analysis
 - Hypothesis driven questions
 - Ex: "are oxidative stress pathways genetically different in individuals with cancer?
 - More common in SNPs than expression
- Agnostic
 - Exploratory analyses across knowledge base
 - Many choices for input
 - Gene-level replication of previously implicated variants
 - Polygenic risk score
 - Ex: "what pathways are these genetic associations aggregating in?"

Mapping SNPs to Genes

- All SNPs in physical proximity of each gene
 - Pros:
 - All/most genes represented
 - Some approaches use LD to help define boundaries

– Cons:

- Varying number of SNPs per gene
- Many of the SNPs may dilute signal
- Defining gene proximity can affect results
- LD is population dependent
 - Need to match LD panel to study population
 - Need raw values or post hoc analysis with summary statistics

Incorporating Functional Info

- eSNPs (Expression associated SNPs)
 - Pros:
 - 1 SNP per gene
 - SNPs functionally associated
 - Cons:
 - Assumes variants effect expression
 - Not all genes have eSNPs
 - eSNPs may be study and tissue dependent
- Hi-C (3D folding)
 - relationship between chromosome organization and genome activity
 - Pros:
 - Helps understand transcription and translation
 - Cons
 - Limited annotation resources
 - Short vs. long range assumptions

Gene summaries

- Initial studies propose different statistics for summarizing the overall gene association prior to enrichment analysis
 - Number/proportion of SNPs with pvalue < 0.05
 - Mean(-log10(pvalue))
 - Min(pvalue) (sentinel SNP)
 - $-1-(1-Min(pvalue))^N$
 - $-1-(1-Min(pvalue))^{(N+1)/2}$

Competitive vs. Self-Contained Tests

Competitive cutoff tests

- Require only permuting SNP or Gene labels
- May only allow to assess relative significance

Self-contained distribution tests

- Require permuting phenotype-genotype relationships
- Resource intensive, may be difficult for large metaanalyses
- Allow to assess overall significance

Competitive vs. Self-Contained Tests

- Self-contained null hypothesis
 - no genes in gene set are differentially expressed

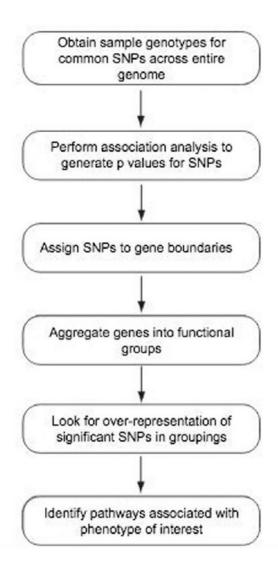
- Competitive null hypothesis
 - genes in gene set are at most as often
 differentially expressed as genes not in gene set

What does this mean for SNP data?

Choice of Pathways/Gene Sets

- Relatively less "signal" in GWAS than in gene expression (GE)
 - GE enrichment typically test which gene sets/pathways show enrichment
 - GWAS enrichment typically test if there is enrichment
- Typically want to be conservative about selecting the number of pathways to test, otherwise will be difficult to overcome multiple testing
- Prioritized Approach:
 - Limited number of specific hypotheses (e.g. gene sets from experiment, co-expression modules, disease-specific pathways/ontologies)
 - Exploratory analyses such as all KEGG/GO sets

Overall Workflow



First approaches: combining p-values

- Compute gene-wise p-value:
 - Select most likely variant 'best' p-value
 - Selected minimum p-value is biased downward
 - Assign 'gene-wise' p-value by permutations (Westfall-Young)
 - Permute samples and compute 'best' p-value for each permutation
 - Compare candidate SNP p-values to this null distribution of 'best' p-values
- Combine p-values by Fisher's method, across SNPs (biased in the presence of correlation)

$$V = -\sum_{g_i \in G} \log(p_i)$$
$$p = P(\chi_{(2k)}^2 > 2V)$$

Next approaches

- Additive model: $\log(\frac{p}{1-p}) = \sum_{g_i \in G} \beta_i n_i$
 - Where n_i indexes the number of allele Bs of a SNP in gene i in the gene set G
 - Select subset of most likely SNP's
 - Fit by logistic regression (glm() in R)
- Significance by permutations
 - Permute sample outcomes
 - Select genes and fit logistic regression again
 - Assess goodness of fit each time
 - Compare observed goodness of fit

Current approaches

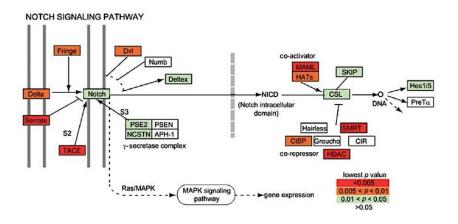
- Adding information about functional annotation to prioritize/select SNPs:
 - ICSNPathway (Zhang et al 2011)
- Use kth best SNP as the representative p-value combined with permutation testing
 - GSA-SNP2 (Nam et al. 2010, Yoon et al 2018)
 - Avoids bias from randomly significant SNPs
 - Loses power if functional SNPs are included

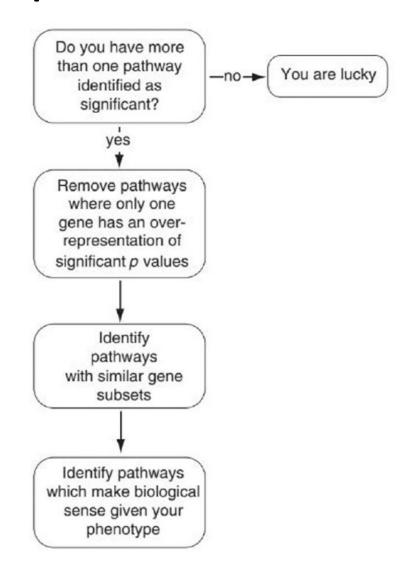
Current approaches

- Using ranked enrichment tests
 - Test the enrichment of a gene or gene set's SNPs at the significant end of a list of ranked SNP p-values
 - VEGAS2 (Liu et al 2010; Mishra & MacGregor 2015;
 Mishra & MacGregor 2017)
 - Accounts for all SNPs
 - Reduces effects of false positive GWAS through enrichment of moderately associate SNPs
 - Doesn't negate strongly significant associations
 - Computationally intensive

Follow-up

- Visualization
- Interpretation





Some Specific Methods

- i-GSEA4GWAS
- MAGENTA/FUMA
 - Meta-Analysis Gene-set Enrichment of variant Associations

i-GSEA4GWAS

- Zhang et al. Nucl Acids Res (2010)
- http://gsea4gwas.psych.ac.cn/
- Categorizes genes as significant or not significant
 - Significant: At least 1 SNP in the top 5% of SNPs
 - Does not adjust for gene size
- Pathway score: k/K
 - k = Proportion of significant genes in the geneset
 - K = Proportion of significant genes in the GWAS
- FDR assessed by permuting SNP labels

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RUN

CLEAR

Improved - Gene Set Enrichment Analysis for Genome-Wide Association Study

A web server for identification of pathways/gene sets associated with traits

☑ Load demo data ②				
Job name: untitled RUN CLEAR	Email (links for result will be sent to your email):			
Select data type: SNP CNV Gene				
GWAS file: Choose File no file selected	-logarithm transformation (necessary ONLY for P-value data)			
elect mapping rules of SNPs->genes 500kb upstream and downstream of gene 20kb upstream and downstream of gene	100kb upstream and downstream of gene			
iii within gene	functional SNP (nonsynonymous, stop gained/lost, frame shift, essential splice site, regulatory region)			
ene set database canonical pathways	GO cellular component			
canonical pathways GO biological process GO molecular function R upload your own gene sets file:	Number of genes in gene set @			
canonical pathways GO biological process GO molecular function R upload your own gene sets file: Choose file no file selected ptions for gene set database Limit gene sets by keyword (e.g. immune). The keyword can be				

Results

Pathway/Gene set name	Description	Manhattan plot 🔮	p. value	FDR	genes/Selected genes/All genes 2	
HSA04950 MATURITY ONSET DIABETES OF THE YOUNG View Detail	Genes involved in ma More	in a second	< 0.001	0.0030		
PROSTAGLANDIN AND LEUKOTRIENE METABOLISM View Detail	More		More \$ 0.001		0.0085	13/27/32
HSA00565 ETHER LIPID METABOLISM View Detail	Genes involved in et More	Macananian	< 0.001	0.0125	15/28/31	
DNA REPAIR View Detail	Genes annotated by t More	d by		0.0135	41/113/125	
NTHIPATHWAY View Detail	Hemophilus influenza More	Introdución	< 0.001	0.0142	12/21/24	
NEGATIVE REGULATION OF DEVELOPMENTAL PROCESS View Detail	Genes annotated by t More	microsomia.	< 0.001	0.014571428	66/175/197	
HSA04330 NOTCH SIGNALING PATHWAY View Detail	Genes involved in No More	microsolenia	< 0.001	0.016	16/35/47	
ENZYME LINKED RECEPTOR PROTEIN SIGNALING PATHWAY	Genes annotated by	Various as	< 0.001	0.020875	60/136/140	

MAGENTA/FUMA

- Segre et al. PLoS Genetics (2010)
- Software download:
 - http://www.broadinstitute.org/mpg/magenta/
 - Requires MATLAB!!
 - Less convenient, but more customizable than iGSEA4GWAS
- Customizable proportion of "significant" genes
- Customizable gene window (upstream & downstream)
- Option for Rank-Sum test
- Gene Summary = min(p)
 - Uses stepwise regression to adjust for multiple possible factors: e.g. gene size, SNP density

MAGENTA Results

*		95% Cutoff (Top 5%)			75% Cutoff (Top 25%)			
GS	NOMINAL GSEA PVAL	A Company of the Comp	The state of the s		NOMINAL GSEA PVAL	and the second s		OBS # GENES
positive regulation of osteoblast differentiation	3.36E-01	8.02E-01	1	2	3.00E-04	7.91E-02	6	14
one-carbon metabolic process	2.20E-03	3.55E-01	1	6	1.60E-03	1.44E-01	7	15
placenta development	3.36E-01	8.06E-01	1	2	4.00E-04	1.45E-01	6	14
carbohydrate transport	8.19E-01	9.46E-01	2	1	3.20E-03	3.45E-01	8	16

FUMA

Watanabe et al, Nature Comm 2017

- Implementation of MAGENTA
- Functional annotation
- Pathway analysis
- Visualization
- •

Other Adaptations of GSEA

- Order log-odds ratios or linkage p-values for all SNPs
- Map SNPs to genes, and genes to groups
- Use linkage p-values in place of t-scores in GSEA
 - Compare distribution of log-odds ratios for SNPs in group to randomly selected SNP's from the chip

Pathway Analysis for Rare Variants

- Low frequency (1% 5% MAF) and rare variants (<1%) require additional considerations
- Off the shelf use of GWAS pathway methods may not be appropriate
 - Generally, rare variants need to be weighted to have any power
 - One and two stage options
 - Using variant level data
 - Collapsing variant level data into genes/regions/pathways

Pathway Analysis for Rare Variants

- Power is highly dependent on how closely the analysis plan matches the true underlying etiology
- Rare variant common disease (RVCD) hypothesis
 - Generally assumed RVs will have high effect sizes and/or direct functional consequences
 - Not always true
 - Ex: Missense mutations can have small effect sizes, with weak selective pressure

Pathway Analysis for Rare Variants

- Example methods:
 - aSPU
 - Pan et al American Journal of Human Genetics 2015
 - Smoothed functional principal components analysis
 - Zhao et al European Journal of Human Genetics 2015
 - Bayesian methods
 - Han et al 2019 bioRxiv doi: https://doi.org/10.1101/828061

Summary Points for GWAS

- In GWAS, few SNPs typically reach genome-wide significance
- Biological function of those that do can take years of work to unravel
- Incorporating biological information (expression, pathways, etc) can help interpret and further explore GWAS results
- Enrichment tests can be used to explore biological pathway enrichment
 - Different tests tell you different things
- Annotation choices very different that in gene expression data, though still rely on the same resources.... not necessarily so for other 'omics"
- Methods for rare variants are evolving

Questions?