Mod 12 Self-Study Material 2: Stata for Windows

Entering data into Stata

1) Enter data directly into Stata.

Open the **Data Editor** window by clicking on the spreadsheet icon on the toolbar. Columns corresponds to variables, rows correspond to observations. Enter the data directly in the spreadsheet then click column names Var1, Var2 to change the variable names. Then close the data editor.

2) Open Stata format data file

Click the **open(use)** icon on the toolbar or use command "**use** filename"

3) Import data from text (ASCII) files.

To open an ASCII (or text) file in Stata, go into the **File** pull-down menu and select **Import**. From the submenu, you can choose to import data created by a spreadsheet, in fixed format, in fixed format with a dictionary, or unformatted. Or use **"infile var1 var2 using filename"** or **"insheet using filename"** for data created by a spreadsheet.

Basic commands

Note: Stata is case-sensitive: small letter not capital letters for commands. Up cases can be used for variable names.

log: to create a log file to store all the used commands and output excluding graphs
cmdlog: to make a record of what you type during your Stata session
#review: to display the last few lines typed at the terminal. For example, #review by itself lists the last 5 lines and "#review 10" would list the last 10 lines.
describe: to describe a dataset
list: to list the contents of a dataset
codebook: to list detailed contents of a dataset
summarize: to calculate and displays a variety of univariate summary statistics
generate: to generate new variable
replace: to change the contents of an existing variable

Save files

To save data set click **Save** icon on the tool bar To save log file, use "log" or click **Begin Log** icon on the toolbar To save graphs, after you create a graph, go to the **File** menu and click **Save Graph**

One can copy selected Stata output and Stata graphs either from **Edit** menu or using Ctrl+c into Word file. For wide table use **Curier New** font with size 8, 9 or 10 so that the table columns will align properly.

Write a do-file

To include a series of commands so that you can repeat the analysis later without typing commands again.

Stata help

Help menu is very useful. You can search contents, State command. Or search internet.

Survival Analysis

Data description: The first data we're going to use in this lab is **myelomatosis data** (Peto et al. 1977), which includes 25 patients diagnosed with myelomatosis. These patients are randomly assigned to two treatments.

DUR: contains the time in days from the point of randomization to either death or censoring.

STATUS: coded 1 if uncensored, 0 if censored.

TREAT: indicates the treatment patients are randomly assigned to.

RENAL: renal functioning at the time of randomization, coded 1 if normal, 0 if impaired.

1). Input data into STATA.

If data file is already in STATA format, you can double click on it to open. The file you download from course website, myel.dta, is the myelomatosis data in STATA format.

2). Declare data to be survival-time data

In order to do survival analysis in STATA, we first need to let STATA know that the data we are dealing with is survival-time data. To declare data to be survival-time data, we use the following command

```
stset dur, failure(status==1)
failure event: status == 1
```

```
obs. time interval: (0, dur]
exit on or before: failure
25 total obs.
0 exclusions
25 obs. remaining, representing
17 failures in single record/single failure data
15334 total analysis time at risk, at risk from t = 0
earliest observed entry t = 0
last observed exit t = 2240
```

When the analysis is done and if you want to go back to normal mode, we can use command stset, clear to make STATA forget that the data are survival-time data.

3). Describe survival-time data

stdes presents a brief description of the survival-time data. It doesn't do any analysis, but only summarizes the information.

Here is the description for myel.dta

Here we set dur to be the time variable and use status to indicate censor (=0) or not (=1). The output of stdes tells us that there are 25 objects at total. We have single record for each object. The table listed mean/minimum/median/maximum for the exit time and time at risk for each object. There are 8 objects censored.

4). Summarize survival-time data

stsum presents summary statistics -- time at risk, incidence rate, number of subjects, and the 25th, 50th, and 75th percentiles of survival time.

•	stsum								
failure _d: status == 1 analysis time _t: dur									
	I		incidence	no. of	Su	rvival time			
		time at risk	rate	subjects	25%	50%	75%		
	total	15334	.0011086	25	63	210			

5). Generate, graph, and list the survivor and cumulative hazard functions

sts graph graphs the estimated survivor or Nelson-Aalen cumulative hazard function.

sts list lists the estimated survivor and related functions.

sts generate creates new variables containing the estimated survivor function, the Nelson-Aalen cumulative hazard function, or related functions.

sts graph /*graphs the estimated survivor function*/



. sts graph, by(treat)





. sts graph, hazard by (treat)



. sts graph, na by(treat)



. sts list, by(treat) $% K-M \ Survival \ For the survival transmission <math display="inline">K-M \ Survival \ Survival$

failure _d: status analysis time _t: dur

	Beg.		Net	Survivor	Std.		
Time	Total	Fail	Lost	Function	Error	[95% Conf	. Int.]
treat=1							
8	12	2	0	0.8333	0.1076	0.4817	0.9555
52	10	1	0	0.7500	0.1250	0.4084	0.9117
63	9	2	0	0.5833	0.1423	0.2701	0.8009
220	7	1	0	0.5000	0.1443	0.2085	0.7361
365	6	0	1	0.5000	0.1443	0.2085	0.7361
852	5	0	1	0.5000	0.1443	0.2085	0.7361
1296	4	0	1	0.5000	0.1443	0.2085	0.7361
1328	3	0	1	0.5000	0.1443	0.2085	0.7361
1460	2	0	1	0.5000	0.1443	0.2085	0.7361
1976	1	0	1	0.5000	0.1443	0.2085	0.7361
treat=2							
13	13	1	0	0.9231	0.0739	0.5664	0.9888
18	12	1	0	0.8462	0.1001	0.5122	0.9591
23	11	1	0	0.7692	0.1169	0.4421	0.9191
70	10	1	0	0.6923	0.1280	0.3734	0.8718
76	9	1	0	0.6154	0.1349	0.3083	0.8184
180	8	1	0	0.5385	0.1383	0.2477	0.7599
195	7	1	0	0.4615	0.1383	0.1916	0.6964
210	6	1	0	0.3846	0.1349	0.1405	0.6280
632	5	1	0	0.3077	0.1280	0.0950	0.5543
700	4	1	0	0.2308	0.1169	0.0558	0.4746
1296	3	1	0	0.1538	0.1001	0.0248	0.3878
1990	2	0	1	0.1538	0.1001	0.0248	0.3878
2240	1	0	1	0.1538	0.1001	0.0248	0.3878

. sts list, by(treat) compare *lists the estimated survivor and related functions.

failure _d: status == 1
analysis time _t: dur

		Survivor	Function
treat		1	2
time	8	0.8333	1.0000
	287	0.5000	0.3846
	566	0.5000	0.3846
	845	0.5000	0.2308
	1124	0.5000	0.2308
	1403	0.5000	0.1538
	1682	0.5000	0.1538
	1961	0.5000	0.1538
	2240		0.1538

sts generate surv=s, by(treat)

creates new variables containing the Kaplan-Meier survivor function. Surv is the name of the new generated variable, which contains Kaplan-Meier survivor function.

. sts gen surv=s, by(treat)

. list surv

						~				
						S	u	r	V	
1.	•	8	3	3	3	3	3	3	3	
2.	•	5	3	8	4	6	1	5	4	
3.		3	0	7	6	9	2	3	1	
4.									5	
5								7	5	
6		1	5	S	Q	л	6	1	5	
· ·	•	Ŧ	J	5	0	4	0	Ŧ	5	
/•		_	_	_	_	_	_	:	5	
8.	•	5	8	3	3	3	3	3	3	
9.	•	4	6	1	5	3	8	4	6	
10.	•	6	1	5	3	8	4	6	2	
11.	•	6	9	2	3	0	7	6	9	
12.		8	3	3	3	3	3	3	3	
13.		9	2	3	0	7	6	9	2	
14		1	5	۔ ع	Ř	Δ	6	1	5	
15	•	-	0	0	0	1	0	-	5	
10.		0	л	c	1	F	\sim	•	5	
10.	•	8	4	0	1	5	3	8	2	
L/.	•	2	3	0	/	6	9	2	3	
18.								•	5	
19.								•	5	
20.	•	3	8	4	6	1	5	3	8	
21.		5	8	3	3	3	3	3	3	
22.									5	
23.		1	5	3	8	4	6	1	5	
24	•	-	Ĵ	Č	Ĵ	-	Ű	1	5	
2- 1. 25		7	c	0	\sim	S	\sim	;	7	
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You can also generate hazard function, Greenwood pointwise standard error, or confidence interval of survival function, by

```
sts generate haz=h, by(treat)
sts generate gse=se, by(treat)
sts generate lowerbound=lb, by(treat)
sts generate upperbound=ub, by(treat)
```

Then plot them using graph command

6. Two Sample Testing in Stata

sts test -- Test equality of survivor functions

sts test varlist [if] [in] [, options]

options description

logrank perform log-rank test of equality; the default

<u>cox</u> perform Cox test of equality

wilcoxon perform Wilcoxon-Breslow-Gehan test of equality (The weights w=n the number of subjects at risk at each interval.)

<u>tware</u> perform Tarone-Ware test of equality (This test is the same as the Wilcoxon test, with the exception that the weight function $w = n^{1/2}$)

peto perform Peto-Prentice test of equality (The only difference between the Wilcoxon test and this one is that the weight function is approximately equal to the K-M survival Function)

 $\mathbf{fh}(p q)$ perform generalized Fleming-Harrington test of equality

trend test trend of the survivor function across three or more ordered groups

strata(varlist) perform stratified test on varlist, displaying overall test results

<u>detail</u> display individual test results; modifies strata()

. sts test treat tests the equality of the survivor function across two different treatment groups.

```
. sts test treat
     failure _d: status == 1
  analysis time _t: dur
Log-rank test for equality of survivor functions
| Events Events
treat | observed expected
    | Events
                 Events
1 | 6 8.34
2 | 11 8.66
_____
Total | 17 17.00
        chi2(1) = 1.31
Pr>chi2 = 0.2519
sts test treat, wilcoxon
  failure _d: status == 1
analysis time _t: dur
Wilcoxon (Breslow) test for equality of survivor functions
| Events Events Sum of
treat | observed expected ranks
| 6 8.34 -18
| 11 8.66 18
1
2
17
Total |
                  17.00
                               0
```

chi2(1) = 0.25 Pr>chi2 = 0.6178 $\ensuremath{\mathtt{P}}\xspace$ value of less than 0.05 is an evidence that there is significant difference between two survival functions.